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(54) Title: PLANT FATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

(57) Abstract

By this invention, compositions and methods of use related to β -ketoacyl-ACP synthase of special interest are synthases obtainable from *Cuphea* species. Amino acid and nucleic acid for synthase protein factors are provided, as well as methods to utilize such sequences in constructs for production of genetically engineered plants having altered fatty acid compositions. Of particular interest is the expression of synthase protein factors in conjunction with expression of plant medium-chain acyl-ACP thioesterases for production of increased levels and/or modified ratios of medium-chain fatty acids in oils of transgenic plant seeds.

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PLANT FATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

INTRODUCTION

Field of Invention

The present invention is directed to genes encoding plant fatty acid synthase enzymes relevant to fatty acid synthesis in plants, and to methods of using such genes in combination with genes encoding plant medium-chain preferring thioesterase proteins. Such uses provide a method to increase the levels of medium-chain fatty acids that may be produced in seed oils of transgenic plants.

Background

Higher plants synthesize fatty acids via a common metabolic pathway. In developing seeds, where fatty acids attached to triglycerides are stored as a source of energy for further germination, the fatty acid synthesis pathway is located in the plastids. The first step is the formation of acetyl-ACP (acyl carrier protein) from acetyl-CoA and ACP catalyzed by a short chain preferring condensing enzyme, ß-ketoacyl-ACP synthase (KAS) III. Elongation of acetyl-ACP to 16- and 18- carbon fatty acids involves the cyclical action of the following sequence of reactions: condensation with a two-carbon unit from malonyl-ACP to form a longer ß-ketoacyl-ACP (ß-ketoacyl-ACP synthase), reduction of the

keto-function to an alcohol (ß-ketoacyl-ACP reductase), dehydration to form an enoyl-ACP (ß-hydroxyacyl-ACP dehydrase), and finally reduction of the enoyl-ACP to form the elongated saturated acyl-ACP (enoyl-ACP reductase). ß-ketoacyl-ACP synthase I (KAS I), is primarily responsible for elongation up to palmitoyl-ACP (C16:0), whereas ß-ketoacyl-ACP synthase II (KAS II) is predominantly responsible for the final elongation to stearoyl-ACP (C18:0).

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Genes encoding peptide components of ß-ketoacyl-ACP synthases I and II have been cloned from a number of higher plant species, including castor (Ricinus communis) and Brassica species (USPN 5,510,255). KAS I activity was associated with a single synthase protein factor having an approximate molecular weight of 50 kD (synthase factor B) and KAS II activity was associated with a combination of two synthase protein factors, the 50 kD synthase factor B and a 46 kd protein designated synthase factor A. Cloning and sequence of a plant gene encoding a KAS III protein has been reported by Tai and Jaworski (Plant Physiol. (1993) 103:1361-1367).

The end products of plant fatty acid synthetase activities are usually 16- and 18-carbon fatty acids. There are, however, several plant families that store large amounts of 8- to 14-carbon (medium-chain) fatty acids in their oilseeds. Recent studies with Umbellularia californica (California bay), a plant that produces seed oil rich in lauric acid (12:0), have demonstrated the existence of a medium-chain-specific isozyme of acyl-ACP thioesterase

in the seed plastids. Subsequent purification of the 12:0-ACP thioesterase from Umbellularia californica led to the cloning of a thioesterase cDNA which was expressed in seeds of Arabidopsis and Brassica resulting in a substantial accumulation of lauric acid in the triglyceride pools of these transgenic seeds (USPN 5,512,482). These results and subsequent studies with medium-chain thioesterases from other plant species have confirmed the chain-length-determining role of acyl-ACP thioesterases during de novo fatty acid biosynthesis (T. Voelker (1996) Genetic Engineering, Ed. J. K. Setlow, Vol. 18, pgs. 111-133).

DESCRIPTION OF THE FIGURES

Figure 1. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor B clone chKAS B-2 are provided. Figure 2. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor B clone chKAS B-31-7 are provided. Figure 3. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor A clone chKAS A-2-7 are provided. Figure 4. DNA and translated amino acid sequence of Cuphea 20 hookeriana KAS factor A clone chKAS A-1-6 are provided. Figure 5. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/7-8 are provided. Figure 6. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/8-7A are provided. 25 Figure 7. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p7-6A are provided. Figure 8. Preliminary DNA sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p8-9A is provided.

- Figure 9. DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 are provided.
 - Figure 10. The activity profile for purified cpuKAS B/8-7A using various acyl-ACP substrates is provided.
- Figure 11. The activity profile for purified chKAS A-2-7 and chKAS A-1-6 using various acyl-ACP substrates is provided.
 - Figure 12. The activity profile for purified castor KAS factor A using various acyl-ACP substrates is provided.
- 10 Figure 13. The activity profile for purified castor KAS factor B using various acyl-ACP substrates is provided.

 Figure 14. A graph showing the number of plants arranged according to C8:0 content for transgenic plants containing CpFatB1 versus transgenic plants containing CpFatB1 + chKAS

A-2-7 is provided.

- Figure 15. Graphs showing the %C10/%C8 ratios in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
- Figure 16. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
- Figure 17. Graphs showing the %C10/%C8 ratios in transgenic
 plants containing ChFatB2 (4804-22-357) and in plants
 resulting from crosses between 4804-22-357 and 5413-17 (chKAS
 A-2-7 + CpFatB1 plants) are provided.
 - Figure 18. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in

plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.

Figure 19. Graphs showing the %C12:0 in transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 20. Graph showing the relative proportions of C12:0 and C14:0 fatty acids in the seeds of transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 21. Graphs showing the %C18:0 in transgenic plants containing Garm FatB1 (5266) and in seeds of plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 22. The activity profile of Ch KAS A in protein extracts from transgenic plants containing Ch KAS A-2-7. Extracts were preptreated with the indicated concentrations of cerulenin.

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SUMMARY OF THE INVENTION

By this invention, compositions and methods of use related to ß-ketoacyl-ACP synthase (KAS) are provided. Also of interest are methods and compositions of amino acid and nucleic acid sequences related to biologically active plant synthase(s).

In particular, genes encoding KAS protein factors A and B from Cuphea species are provided. The KAS genes are of interest for use in a variety of applications, and may be

used to provide synthase I and/or synthase II activities in transformed host cells, including bacterial cells, such as E. coli, and plant cells. Synthase activities are distinguished by the preferential activity towards longer and shorter acyl-ACPs as well as by the sensitivity towards the KAS specific inhibitor, cerulenin. Synthase protein preparations having preferential activity towards medium chain length acyl-ACPs are synthase I-type or KAS I. The KAS I class is sensitive to inhibition by cerulenin at concentrations as low as $1\mu M$. Synthases having preferential activity towards longer chain length acyl-ACPs are synthase II-type or KAS II. The KAS enzymes of the II-type are also sensitive to cerulenin, but at higher concentrations (50 μM). Synthase III-type enzymes have preferential activity towards short chain length acyl-ACPs and are insensitive to cerulenin inhibition.

Nucleic acid sequences encoding a synthase protein may be employed in nucleic acid constructs to modulate the amount of synthase activity present in the host cell, especially the relative amounts of synthase I-type, synthase II-type and synthase III-type activity when the host cell is a plant host cell. In addition, nucleic acid constructs may be designed to decrease expression of endogenous synthase in a plant cell as well. One example is the use of an antisense synthase sequence under the control of a promoter capable of expression in at least those plant cells which normally produce the enzyme.

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Of particular interest in the present invention is the coordinate expression of a synthase protein with the

expression of thioesterase proteins. For example, coordinated expression of synthase factor A and a medium-chain thioesterase provides a method for increasing the level of medium-chain fatty acids which may be harvested from transgenic plant seeds. Furthermore, coordinated expression of a synthase factor A gene with plant medium-chain thioesterase proteins also provides a method by which the ratios of various medium-chain fatty acids produced in a transgenic plant may be modified. For example, by expression of a synthase factor A, it is possible to increase the ratio of C10/C8 fatty acids which are produced in plant seed oils as the result of expression of a thioesterase having activity on C8 and C10 fatty acids.

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DETAILED DESCRIPTION OF THE INVENTION

A plant synthase factor protein of this invention includes a sequence of amino acids or polypeptide which is required for catalyzation of a condensation reaction between an acyl-ACP having a chain length of C2 to C16 and malonyl-ACP in a plant host cell. A particular plant synthase factor protein may be capable of catalyzing a synthase reaction in a plant host cell (for example as a monomer or homodimer) or may be one component of a multiple peptide enzyme which is capable of catalyzing a synthase reaction in a plant host cell, i.e. one peptide of a heterodimer.

Synthase I (KAS I) demonstrates preferential activity towards acyl-ACPs having shorter carbon chains, C2-C14 and is sensitive to inhibition by cerulenin at concentrations of 1µM. Synthase II (KAS II) demonstrates preferential

activity towards acyl-ACPs having longer carbon chains, C14-C16, and is inhibited by concentrations of cerulenin (50µM). Synthase III demonstrates preferential activity towards acyl-CoAs having very short carbon chains, C2 to C6, and is insensitive to inhibition by cerulenin.

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Synthase factors A and B, and synthase III proteins obtained from medium-chain fatty acid producing plant species of the genus Cuphea are described herein. As described in the following Examples, synthase A from C. hookeriana is naturally expressed at a high level and only in the seeds. C. hookeriana synthase B is expressed at low levels in all tissues examined. Expression of synthase A and synthase B factors in E. coli and purification of the resulting proteins is employed to determine activity of the various synthase factors. Results of these analyses indicate that synthase factor A from Cuphea hookeriana has the greatest activity on 6:0-ACP substrates, whereas synthase factor B from Cuphea pullcherrima has greatest activity on 14:0-ACP. Similar studies with synthase factors A and B from castor demonstrate similar activity profiles between the factor B synthase proteins from Cuphea and The synthase A clone from castor, however, castor. demonstrates a preference for 14:0-ACP substrate.

Expression of a Cuphea hookeriana KAS A protein in transgenic plant seeds which normally do not produce medium-chain fatty acids does not result in any detectable modification of the fatty acid types and contents produced in such seeds. However, when Cuphea hookeriana KAS A protein is expressed in conjunction with expression of a

medium-chain acyl-ACP thioesterase capable of providing for production of C8 and C10 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are observed. In addition, where significant amounts of C8 and C10 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C8 and C10 fatty acids that are obtained. For example, an increased proportion of C10 fatty acids may be obtained by co-expression of Cuphea hookeriana ChFatB2 thioesterase and a chKAS A synthase factor proteins.

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Furthermore, when Cuphea hookeriana KAS A protein is expressed in conjunction with expression of a medium-chain acyl-ACP thioesterase capable of providing for production of C12 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are also observed. In addition, where significant amounts of C12 and C14 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C12 and C14 fatty acids that are obtained. For example, an increased proportion of C12 fatty acids may be obtained by co-expression of UC FatB1 thioesterase and a chKAS A synthase factor proteins.

However, when Cuphea hookeriana KAS A protein is expressed in conjunction with the expression of a long-chain

acyl-ACP thioesterase capable of providing for production of C18 and C18:1 fatty acids in plant seed oils, no effect on the production of long chain fatty acids was observed.

Furthermore, when plants transformed to express a long chain acyl-ACP thioesterase from mangosteen (GarmFatA1, U.S. Patent Application No. 08/440,845), which preferentially hydrolyzes C18:0 and C18:1 fatty acyl-ACPs, are crossed with nontransformed control plants, a significant reduction in the levels of C18:0 is obtained. Similar reductions are also observed in the levels of C18:0 in the seeds of plants resulting from crosses between plants transformed to express the GarmFatA1 and plants expressing the Cuphea hookeriana KAS A protein.

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Thus, the instant invention provides methods of increasing and/or altering the medium-chain fatty acid compositions in transgenic plant seed oils by co-expression of medium-chain acyl-ACP thioesterases with synthase factor proteins. Furthermore, various combinations of synthase factors and medium-chain thioesterases may be achieved depending upon the particular fatty acids desired. For example, for increased production of C14 fatty acids, synthase protein factors may be expressed in combination with a C14 thioesterase, for example from Cuphea palustris or nutmeg may be employed (WO 96/23892). In addition, thioesterase expression may be combined with a number of different synthase factor proteins for additional effects on medium-chain fatty acid composition.

Synthases of use in the present invention include modified amino acid sequences, such as sequences which have

been mutated, truncated, increased and the like, as well as such sequences which are partially or wholly artificially synthesized. The synthase protein encoding sequences provided herein may be employed in probes for further screening or used in genetic engineering constructs for transcription or transcription and translation in host cells, especially plant host cells. One skilled in the art will readily recognize that antibody preparations, nucleic acid probes (DNA and RNA) and the like may be prepared and used to screen and recover synthases and/or synthase nucleic acid sequences from other sources. Typically, a homologously related nucleic acid sequence will show at least about 60% homology, and more preferably at least about 70% homology, between the R. communis synthase and the given plant synthase of interest, excluding any deletions which may be present. Homology is determined upon comparison of sequence information, nucleic acid or amino acid, or through hybridization reactions.

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Recombinant constructs containing a nucleic acid sequence encoding a synthase protein factor or nucleic acid sequences encoding a synthase protein factor and a medium-chain acyl-ACP thioesterase may be prepared by methods well known in the art. Constructs may be designed to produce synthase in either prokaryotic or eukaryotic cells. The increased expression of a synthase in a plant cell, particularly in conjunction with expression of medium-chain thioesterases, or decreasing the amount of endogenous synthase observed in plant cells are of special interest.

Synthase protein factors may be used, alone or in combination, to catalyze the elongating condensation reactions of fatty acid synthesis depending upon the desired result. For example, rate influencing synthase activity may reside in synthase I-type, synthase II-type, synthase III-type or in a combination of these enzymes. Furthermore, synthase activities may rely on a combination of the various synthase factors described herein.

Constructs which contain elements to provide the transcription and translation of a nucleic acid sequence of interest in a host cell are "expression cassettes".

Depending upon the host, the regulatory regions will vary, including regions from structural genes from viruses, plasmid or chromosomal genes, or the like. For expression in prokaryotic or eukaryotic microorganisms, particularly unicellular hosts, a wide variety of constitutive or regulatable promoters may be employed. Among transcriptional initiation regions which have been described are regions from bacterial and yeast hosts, such as E. coli, B. subtilis, Saccharomyces cerevisiae, including genes such as &-galactosidase, T7 polymerase, trp-lac (tac), trp E and the like.

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An expression cassette for expression of synthase in a plant cell will include, in the 5' to 3' direction of transcription, a transcription and translation initiation control regulatory region (also known as a "promoter") functional in a plant cell, a nucleic acid sequence encoding a synthase, and a transcription termination region.

Numerous transcription initiation regions are available

which provide for a wide variety of constitutive or regulatable, e.g., inducible, transcription of the desaturase structural gene. Among transcriptional initiation regions used for plants are such regions associated with cauliflower mosaic viruses (35S, 19S), and structural genes such as for nopaline synthase or mannopine synthase or napin and ACP promoters, etc. The transcription/ translation initiation regions corresponding to such structural genes are found immediately 5' upstream to the respective start codons. Thus, depending upon the intended use, different promoters may be desired.

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Of special interest in this invention are the use of promoters which are capable of preferentially expressing the synthase in seed tissue, in particular, at early stages of seed oil formation. Examples of such seed-specific promoters include the region immediately 5' upstream of a napin or seed ACP genes such as described in USPN 5,420,034, desaturase genes such as described in Thompson et al (Proc. Nat. Acad. Sci. (1991) 88:2578-2582), or a Bce-4 gene such as described in USPN 5,530,194. Alternatively, the use of the 5' regulatory region associated with the plant synthase structural gene, i.e., the region immediately 5' upstream to a plant synthase structural gene and/or the transcription termination regions found immediately 3' downstream to the plant synthase structural gene, may often be desired. general, promoters will be selected based upon their expression profile which may change given the particular application.

In addition, one may choose to provide for the transcription or transcription and translation of one or more other sequences of interest in concert with the expression or anti-sense of the synthase sequence, particularly medium-chain plant thioesterases such as described in USPN 5,512,482, to affect alterations in the amounts and/or composition of plant oils.

When one wishes to provide a plant transformed for the combined effect of more than one nucleic acid sequence of interest, a separate nucleic acid construct may be provided for each or the constructs may both be present on the same plant transformation construct. The constructs may be introduced into the host cells by the same or different methods, including the introduction of such a trait by crossing transgenic plants via traditional plant breeding methods, so long as the resulting product is a plant having both characteristics integrated into its genome.

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Normally, included with the DNA construct will be a structural gene having the necessary regulatory regions for expression in a host and providing for selection of transformed cells. The gene may provide for resistance to a cytotoxic agent, e.g. antibiotic, heavy metal, toxin, etc., complementation providing prototrophy to an auxotrophic host, viral immunity or the like. Depending upon the number of different host species into which the expression construct or components thereof are introduced, one or more markers may be employed, where different conditions for selection are used for the different hosts.

The manner in which the DNA construct is introduced into the plant host is not critical to this invention. Any method which provides for efficient transformation may be employed. Various methods for plant cell transformation include the use of Ti- or Ri-plasmids, microinjection, electroporation, liposome fusion, DNA bombardment or the like. In many instances, it will be desirable to have the construct bordered on one or both sides by T-DNA, particularly having the left and right borders, more particularly the right border. This is particularly useful when the construct uses A. tumefaciens or A. rhizogenes as a mode for transformation, although the T-DNA borders may find use with other modes of transformation.

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The expression constructs may be employed with a wide variety of plant life, particularly plant life involved in the production of vegetable oils. These plants include, but are not limited to rapeseed, peanut, sunflower, safflower, cotton, soybean, corn and oilseed palm.

For transformation of plant cells using Agrobacterium, explants may be combined and incubated with the transformed Agrobacterium for sufficient time for transformation, the bacteria killed, and the plant cells cultured in an appropriate selective medium. Once callus forms, shoot formation can be encouraged by employing the appropriate plant hormones in accordance with known methods and the shoots transferred to rooting medium for regeneration of plants. The plants may then be grown to seed and the seed used to establish repetitive generations and for isolation of vegetable oils.

The invention now being generally described, it will be more readily understood by reference to the following examples which are included for purposes of illustration only and are not intended to limit the present invention.

EXAMPLES

Example 1 Cuphea KAS Factor A and B Gene Cloning

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Total RNA isolated from developing seeds of Cuphea hookeriana and Cuphea pullcherrima was used for cDNA synthesis in commercial 1-based cloning vectors. For cloning each type of KAS gene, approximately 400,000-500,000 unamplified recombinant phage were plated and the plaques transferred to nitrocellulose. For KAS factor B cloning from C. hookeriana, a mixed probe containing Brassica napus KAS factor B and Ricinus communis (Castor) KAS factor B radiolabeled cDNA's was used. Similarly, a mixed probe containing Brassica napus KAS factor A and Ricinus communis KAS factor A cDNA clones was used to obtain C. hookeriana KAS factor A genes. For KASIII, a spinach KASIII cDNA clone obtained from Dr. Jan Jaworski was radiolabeled and used as a probe to isolate a KASIII clone from C. hookeriana. For KAS B and KAS A cloning from C. pullcherrima, C. hookeriana KAS B and KAS A genes chKAS B-2 and chKAS A-2-7 (see below) were radiolabeled and used as probes.

DNA sequence and translated amino acid sequence for Cuphea KAS clones are provided in Figures 1-9. Cuphea hookeriana KAS factor B clones chKAS B-2 and chKAS B-31-7

are provided in Figures 1 and 2. Neither of the clones is full length. Cuphea hookeriana KAS Factor A clones chKAS A-2-7 and chKAS A-1-6 are provided in Figures 3 and 4. chKAS A-2-7 contains the entire encoding sequence for the KAS factor protein. Based on comparison with other plant synthase proteins, the transit peptide is believed to be represented in the amino acids encoded by nucleotides 125-466. chKAS A-1-6 is not a full length clone although some transit peptide encoding sequence is present. Nucleotides 1-180 represent transit peptide encoding sequence, and the mature protein encoding sequence is believed to begin at nucleotide 181.

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Cuphea pullcherrima KAS factor B clones cpuKAS B/7-8 and cpuKAS B/8-7A are provided in Figures 5 and 6. Both of the clones contain the entire encoding sequences for the KAS factor B proteins. The first 35 amino acids of cpuKAS B/7-8 are believed to represent the transit peptide, with the mature protein encoding sequence beginning at nucleotide The first 39 amino acids of cpuKAS B/8-7A are believed 233. to represent the transit peptide, with the mature protein encoding sequence beginning at nucleotide 209. Cuphea pullcherrima KAS factor A clones cpuKAS A/p7-6A and cpuKAS A-p8-9A are provided in Figures 7 and 8. Both of the clones contain the entire encoding sequences for the KAS factor A proteins. Translated amino acid sequence of cpuKAS A/p7-6A The mature protein is believed to begin at the is provided. lysine residue encoded 595-597, and the first 126 amino acids are believed to represent the transit peptide. DNA sequence of KAS A clone cpuKAS A-p8-9A is preliminary.

Further analysis will be conducted to determine final DNA sequence and reveal the amino acid sequence encoded by this gene.

DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 is provided in Figure 9. The encoding sequence from nucleotides 37-144 of chKASIII-27 are believed to encode a transit peptide, and the presumed mature protein encoding sequence is from nucleotides 145-1233.

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Deduced amino acid sequence of the C. hookeriana KAS factor B and KAS factor A cDNA's reveals strong homology to the Brassica napus and Ricinus communis clones previously reported. The C. hookeriana KAS factor B clone is more homologous to the Ricinus and Brassica KAS factor B clones (94% and 91% respectively) than it is to the Ricinus and Brassica KAS factor A clones (60% for both). Furthermore, the C. hookeriana KAS factor A clone is more homologous to the Ricinus and Brassica KAS factor A clones (85% and 82% respectively) than it is the Ricinus and Brassica KAS factor B clone (60% for both). The C. hookeriana KAS factor B cDNAs designated as chKAS B-2 and chKAS B-31-7 are 96% identical within the mature portion of the polypeptide. Similarly, the deduced amino acid sequence of the mature protein regions of the C. hookeriana KAS factor A clones chKAS A-2-7 and chKAS A-1-6 are 96% identical. pullcherrima KAS clones also demonstrate homology to the R. communis and Brassica napus KAS clones. The mature protein portion of all of the KAS factor A family members in the different Cuphea species are 95% identical. Similarly the

mature protein portion of the KAS factor B genes in Cuphea are also 95-97% identical with each other. However there is only approximately 60% sequence identity between KAS factor B and KAS factor A clones either within the same or different species of Cuphea.

Example 2 Levels and Patterns of Expression

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To examine tissue specificity of KAS expression in Cuphea hookeriana, Northern blot analysis was conducted using total RNA isolated from seed, root, leaf and flower tissue. Two separate but identical blots were hybridized with either chKAS B-31-7 or chKAS A-2-7 coding region probes. The data from this RNA blot analysis indicate that KAS B is expressed at a similar level in all tissues examined, whereas KAS A expression is detected only in the These results also demonstrate a different level of seed. expression for each of the synthases. KAS A is an abundant message, whereas KAS B is expressed at low levels. Furthermore, even under highly stringent hybridization conditions (65_C, 0.1 X SSC, 0.5% SDS), the KAS A probe hybridizes equally well with two seed transcripts of 2.3 and The larger hybridizing band is likely the 1.9 kb. transcript of the KAS A-2-7 gene since the size of its cDNA is 2046bp, and the number of clones obtained from cDNA screening corresponds well with the apparent mobility of the mRNA and its abundance on the blot.

Example 3 Expression of Plant KAS Genes in E.coli

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DNA fragments encoding the mature polypeptide of the Cuphea hookeriana KAS A cDNAs and the Cuphea pullcherrima 5 KAS B cDNAs were obtained by PCR and cloned into a QIAexpress expression vector (Qiagene). Experimental conditions for maximum level of expression were determined for all of these clones and the parameters for highest level of soluble fraction were identified. Cells are grown in ECLB media containing 1M sorbitol and 2.5 mM betaine overnight and subcultured as a 1:4 dilution in the same medium. Cells are then grown for 2 hours (to approximately .6-.8 O.D.) and induced with 0.4 mM IPTG and allowed to grow for 5 more hours.

Enzyme activity of the affinity purified recombinant enzymes obtained from over-expression of the chKAS A-2-7 and cpuKAS B/8-7A clones was measured using a wide range of acyl-ACP substrates (6:0- to 16:1-ACP). The activity profile for cpuKAS B/8-7A is provided in Fig.10. demonstrate that the enzyme is active with all acyl-ACP substrates examined, although activity on 6:0 to 14:0-ACP substrates is substantially greater than the activity on 16:0 and 16:1 substrates.

The activity profile of the C. hookeriana KAS A clones chKAS A-2-7 and chKAS A-1-6 is provided in Figure 11. The C. 25 hookeriana KAS A clones are most active with C:6, and have the least activity with C:16:0 substrates. However, the activity of this clone on even the preferred C6:0 substrate

is 50 fold lower than the activity of the *C. pullcherrima* KAS B clones.

A fragment containing the mature protein encoding portion of a R. communis KAS factor A clone was also cloned into a QIAexpress expression vector, expressed in E. coli and the enzyme affinity purified as described above. The activity profile for castor KAS A is provided in Figure 12. Highest activity is observed with C14:0 substrates, although some activity is also seen with C6:0 and C16:1. In comparison, the activity profile obtained from purified R. communis KAS factor B also using the QIAexpress expression system is provided in Figure 13. The KAS B clone demonstrates substantially higher levels of activity (10 fold and higher) than the R. communis KAS A clone. The preference of the KAS factor B for 6:0- to 14:0-ACP substrates is consistent with the previous observations that this protein provides KAS I activity.

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Example 4 KAS and TE Expression in Transgenic Seed

Both the CpFatB1 (*C. hookeriana* thioesterase cDNA;

Dehesh et al. (1996) Plant Physiol. 110:203-210) and the chKAS A-2-7 were PCR amplified, sequenced, and cloned into a napin expression cassette. The napin/cp FatB1 and the napin/KAS A-2-7 fusions were ligated separately into the binary vector pCGN1558 (McBride and Summerfelt (Pl.Mol.Biol. (1990) 14:269-276) and transformed into A. tumefaciens, EHA101. The resulting CpFatB1 binary construct is pCGN5400 and the chKAS A-2-7 construct is pCGN5401. Agrobacterium mediated transformation of a Brassica napus canola variety

was carried out as described by Radke et al. (Theor. Appl. Genet. (1988) 75:685-694; Plant Cell Reports (1992) 11:499-505). Several transgenic events were produced for each of the pCGN5400 and pCGN5401 constructs.

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A double gene construct containing a napin/cpFatB1 expression construct in combination with a napin/chKAS A-2-7 expression construct was also assembled, ligated into a binary vector and used for co-cultivation of a canola Brassica variety. The binary construct containing the chFatB1 and chKAS A-2-7 expression constructs is pCGN5413.

Fatty acid analysis of 26 transgenic lines containing chKAS A-2-7 (5401 lines) showed no significant changes in the oil content or profile as compared to similar analyses of wild type canola seeds of the transformed variety.

Fatty acid analysis of 36 transgenic lines containing cpFatB1 (5400 lines) showed increased levels of C:8 and C:10 in transgenic seeds. The highest level of C:8 observed in a pool seed sample was 4.2 mol%. The C:10 levels were between 30 and 35% of the C:8 content. Fatty acid analysis of 25 transgenic lines containing the TE/KAS A tandem (5413 lines) demonstrated an overall increase in both C:8 and C:10 levels relative to those observed with TE containing lines (5400) alone. In lines containing the cpFatB1 construct alone, the average level of C:8 average were 1.5 mol%, whereas the C:8 average levels in TE/KAS A tandem containing lines was 2.37 mol%. The ratio of C:8 to C:10 remained constant in both populations. The number of transgenic events relative to the C:8 content are presented in Figure 14. These data show that the transgenic events with tandem TE/KAS A construct

yield more lines with higher levels of C:8 than those events with single TE construct. For example, several lines containing nearly 7 mole% C8 were obtained with the TE/KAS A pCGN5413 construct, whereas the highest C8 containing line from the pCGN5400 TE alone transformation contained 4.2 mole% C8.

Half seed analysis of the T3 generation of transgenic canola plants expressing a ChFatB2 (*C. hookeriana* thioesterase; Dehesh et al. (1996) The Plant Journal 9:167-172) indicate that these plant can accumulate up to 22 weight% (33 mol%) of 8:0 and 10:0 fatty acids (4804-22-357). Segregation analysis shows that these transformants contain two loci and that they are now homozygous. Selected plants grown from these half seeds were transferred into the greenhouse and later crossed with T1 transformants that had been transformed with either Cuphea hookeriana KAS A (5401) alone or KAS A/CpFatB1 double constructs (5413).

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Fatty acid analysis of several events resulting from the crosses between transgenic lines containing ChFatB2 (4804-22-357) and chKAS A-2-7 (5401-9), reveal an increase in the ratio of C:10/C:8 levels (Figure 15). This C:10/C:8 ratio in nearly all of the transgenic events containing ChFatB2 TE alone fluctuates between 3 and 6, whereas in the F1 generation of transgenic containing both the TE and the KAS A-2-7, the ratio can be as high as 22. This increase in C:10 levels is accompanied by an increase in the total C:8 and C:10 content (Figure 16). The sum of the C:8 and C:10 fatty acids in the heterozygous F1 lines is as high as those in the homozygous parent line (4804-22-357), whereas the

heterozygous lines usually contain substantially less C:8 and C:10 than the homozygous lines.

Similar results were observed in F1 generation seeds resulting from crosses performed between 4804-22-357 (ChFatB2) and the 5413-17 event (CpFatB1 and chKAS A-2-7 tandem). Levels of C:8 and C:10 in the 5413-17 line were 6.3 and 2.8 mol% respectively. Data presented in Figure 17 show that there is shift towards C:10 fatty acids as was observed with the 4804-22-357 (ChFatB2) x 5401-9 (chKAS A-2-7) crosses. Furthermore, Figure 18 indicates the presence 10 of two separate populations of heterozygotes. containing approximately 9-11 weight percent C:10 + C:8 are believed to represent offspring containing a single copy of the ChFatB1 TE gene and no copies of the CpFatB1 and chKAS A genes from 5413. Those plants containing approximately 15-15 20 weight percent C:10 + C:8 are believed to represent the heterozygotes containing a single ChFatB1 TE gene as well as the CpFatB1 and chKAS A genes from 5413. Thus, the level of the C:10 + C:8 fatty acids does not decrease to 50% of that detected in parent lines when a copy of the ChKAS A gene is 20 present.

To further characterize the chain length specificity of the Cuphea hookeriana KAS A enzyme, crosses between transgenic Brassica napus lines containing a California Bay (Umbellularia californica) 12:0 specific thioesterase, Uc FatB1 (USPN 5,344,771) and chKAS A-2-7 (5401-9) were made. Half seed analysis of transgenic plants containing Uc fatB1 have previuosly indicated that these plants can accumulate up to 52 mol% C12:0 in the seed oil of homozygous dihaploid

lines (LA86DH186). Crosses between the line LA86DH186 and untransformed control *Brassica* demonstrated a decrease in the C12:0 levels.

however, crosses between LA86DH186 and the 5401-9

hemizygous line led to an accumulation of up to 57 mol%

C12:0 in the seed oil of F1 progeny (Figure 19).

Interestingly, in crosses with LA86DH186 x untransformed

control line and LA86DH186 x 5401-9, levels of C14:0 in the

seeds of the F1 progeny decreased to 50% of the levels

obtained in homozygous LA86DH186 lines (Figure 20).

Furthermore, increases in the proportion of C12:0 fatty acid

resulted in a substantial decline in the proportions of all

the long-chain fatty acyl groups (C16:0, C18:0, C18:2, and

C18:3). These results indicate that the ChKAS A-2-7 is an

enzyme with substrate specificity ranging from C6:0 to

C10:0-ACP, and that its over-expression ultimately reduces

the longer chain acyl-ACP pools.

Further evidence is obtained in support of the chain length specificity of the ChKAS A-2-7 in crosses of the 5401-9 line with a transgenic line (5266) expressing an 18:1/18:0 TE from Garcinia mangostana (GarmFatA1, US patent application No. 08/440,845). Transgenic Brassica line 5266 has been shown to accumulate up to 24 mol% C18:0 in the seed oil of homozygous lines (Figure 21). However, in the seed oil of F1 progeny of crosses between 5266 and 5401-9 levels of C18:0 were reduced to approximately 12 mol%. Furthermore, levels of C16:0 generated from these crosses was similar to the levels obtained from the seed oil of nontransgenic control plants.

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Example 5 In vitro Analysis of Plant KAS Enzymes

Seed extracts were prepared from developing seeds of nontransgenic controls or transgenic Brassica expressing chKAS A-2-7 as described in Slabaugh et al. (Plant Journal, 1998 in press) and Leonard et al. (Plant Journal, 1998, in press). In vitro fatty acid synthesis assays were performed as described by Post-Beittenmiller (J. Biol. Chem. (1991), 266:1858-1865). Extracts were concentrated by ammonium sulfate precipitation and desalting using P-6 columns (Bio-Rad, Hercules, CA). Reactions (65 μ 1) contained 0.1M 10 Tris/HCl (pH 8.0), 1 mM dithiothreitol, 25 mM recombinant spinach ACP1, 1 mM NADH, 2 mM NADPH, 50 µM malonyl-CoA, 10 μM [1-14C]acetyl-CoA (50 mCi/mmol), 1mg/ml BSA, and 0.25 mg/ml seed protein. Selected seed extracts were preincubated with cerulenin at 23°C for 10 min. Reaction 15 products were separated on an 18% acrlamide gel containing 2.25M urea, electroblotted onto to nitrocellulose and quntitated by phosporimaging using Image QuaNT software (Molecular Dynamics, Sunnyvale, CA). Authentic acyl-ACPs were run in parallel, immunoblotted and finally detected by 20 anti-ACP serum to confirm fatty acid chain lengths.

The results (Figure 22) indicate that the fatty acid synthesis capabilities of transgenic Brasica (5401-9) seed extracts was greater than that obtained from in the nontransgenic controls as measured by the relative abundance of C8:0- and C10:0-ACP at all time points tested. In addition, pretreatment of the extracts with cerulenin, markedly reduced the synthesis of longer chain fatty acids in both the transgenic and nontransgenic control seed

extracts. However, the extension of the spinach-ACP was much less inhibited in the seed extracts from the transgenic lines than in the seed extracts of nontransgenic control Brassica.

These data further support that Ch KAS A-2-7 is a condensing enzyme active on medium chain acyl-ACPs, and that expression of this enzyme in plants results in enlarged substrate pools to be hydrolyzed by medium-chain specific thioesterases. Furthermore, these data suggest that chKAS A-2-7 also is a cerulenin-resistant condensing enzyme.

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All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claim.

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MISSING UPON TIME OF PUBLICATION

- 13. The construct of Claim 5 wherein said encoding sequence is cpuKAS A/p8-9A.
- 14. The construct of Claim 5 wherein said encoding sequence is chKASIII-27.
- 15. An improved method for producing medium-chain fatty acids in transgenic plant seeds by expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant,

the improvement comprising expression of a plant synthase

factor protein heterologous to said transgenic plant in

conjunction with expression of said plant medium-chain

thioesterase, whereby the percentage of medium-chain fatty

acids produced in seeds expressing both a plant synthase factor

protein and a plant medium-chain thioesterase protein is

increased as compared to the percentage of medium-chain fatty

acids produced in seeds expressing only said plant medium-chain

thioesterase protein.

- 16. The method of Claim 15 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 20 17. The method of Claim 15 wherein said medium-chain thioesterase protein is a CpFatB1 protein.
 - 18. The method of Claim 15 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
- 19. The method of Claim 15 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
 - 20. The method of Claim 19 wherein said synthase factor A protein is from a Cuphea species.

21. The method of Claim 20 wherein said Cuphea species is C. hookeriana or C. pullcherrima.

22. A method of altering the medium-chain fatty acid composition in plant seeds expressing a heterologous plant medium-chain preferring thioesterase, wherein said method comprises

providing for expression of a plant synthase factor protein heterologous to said transgenic plant in conjunction with expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant, whereby the composition of medium-chain fatty acids produced in said seeds is modified as compared to the composition of medium-chain fatty acids produced in seeds expressing said plant medium-chain thioesterase protein in the absence of expression of said plant synthase factor protein.

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- 23. The method of Claim 22 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 24. The method of Claim 22 wherein said medium-chain thioesterase protein is a CpFatBl protein.
- 25. The method of Claim 22 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
 - 26. The method of Claim 22 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
 - 27. The method of Claim 26 wherein said synthase factor A protein is from a Cuphea species.
 - 28. The method of Claim 27 wherein said Cuphea species is C. hookeriana or C. pullcherrima.

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29. The method of Claim 22 wherein said fatty acid composition is enriched for C10 fatty acids.

- 30. The method of Claim 22 wherein said fatty acid composition is enriched for C12 fatty acids.
- 31. The method of Claim 22 wherein said fatty acid composition is enriched for at least one medium chain fatty acid and at least one other medium chain fatty acid is decreased.
- 32. The method of Claim 31 wherein said enriched fatty 10 acid is C12 and said decreased fatty acid is C14.

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48	96	144	192	240	288	336	384
		**1	• •				
GGC Gly	AAG Lys	GGT Gly	CAC	GGG G1y	TCA Ser	GCT Ala	ACT Thr
CCG (Pro)	TCC	GGT Gly	GGT Gly	ATG Met	TAT Tyr	GCC Ala	66C G1y
CCC (Pro	CTC	ATG	AAG Lys	AAC Asn	AAC Asn	GCT Ala	GGA G1y
GAT (Asp	CGC	GGA	GAG Glu	ACA Thr	CCA Pro	CAT	GCT Ala
GTG (GAC	ACA Thr	ATC Ile	ATT Ile	66C 61y	TTC	ATT Ile
CTA (Leu	GCC	GGA G1y	CTT Leu	GCC Ala	ATG Met	TGC Cys	ATG Met
GAA	GGT Gly	GTC Val	TCT Ser	$\mathtt{TAT}\\\mathtt{Ty}_{\mathcal{I}}$	CTC Leu	TAC	CTT Leu
CTA (CTC	CTG Leu	CAG Gln	CCC Pro	GGT Gly	AAC Asn	GAT Asp
GCT	GAT Asp	GTG Val	GTT Val	ATC Ile	TTT Phe	TCC	GCT Ala
GCC	GCC Ala	GGA G1y	GGG Gly	TTC Phe	GAA Glu	ACT Thr	GAG Glu
GCG	CGA Arg	GCC Ala	GAC Asp	TTC	ATC Ile	GCC Ala	GGT Gly
GTG Val	GCA Ala	AGA Arg	TCT Ser	CCT	GCT Ala	TGT Cys	CGT Arg
GCG Ala	TCG Ser	GAG Glu	TTC Phe	ACC Thr	CTC	GCA Ala	CGC
ACC Thr	AAT Asn	AAG Lys	GTC Val	ATC Ile	CTG Leu	ACT Thr	ATC Ile
TCC	AGG Arg	GAC Asp	ACT Thr	AAA Lys	GCC Ala	TCC	CAT
AGC	TGC	ATC Ile	CTG	CGG Arg	TCT Ser	ATT Ile	AAT Asn

IGURE 1 1 OF 4

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454	480	528	576	624	672	720	768	
						:	•	
AGG Arg	TGG Trp	TTG	ATT Ile	ACT Thr	AGC	GCT Ala	ATC Ile	
GCT TGC Ala Cys A	CCC Pro	GTG Val	ATT Ile	ATG Met	AGT	AAT	GCC	
GCT Ala	AGG Arg	GGA Gly	CCG	CAC	GAG	ATA	AAT Asn	
GTG Val	TCT Ser	GCT Ala	GCA Ala	TAT Tyr	ATT Ile	TAC	ATA Ile	
TTT Phe	GCC Ala	GGT Gly	GGA	GCT Ala	TGC	AAT Asn	GAG Glu	
GGC TTT Gly Phe	ACT Thr	GAA Glu	CGA	GAT Asp	TCT	GTC Val	GCC Ala	
GGA Gly	CAG	GGT Gly	AGA Arg	TGT Cys	TCT Ser	GAG Glu	SAT CTC ASP Leu FIGURE 1 2 OF 4	
TTG Leu	CCG Pro	ATG Met	ATG Met	AAC Asn	GTC	GAA Glu	<i>-</i> 7	
666 61y	GAC Asp	GTG Val	GCA	ATC Ile	GGT Gly	CCT	GGG G1y	
ATT Ile	GAT Asp	$ ext{TTT}$	CAT His	GCA Ala	CTT Leu	TCA Ser	GCT	
CCA Pro	AAC Asn	GGT Gly	GAA Glu	GGT Gly	GGT	GTC	CTA Leu	
ATT Ile	AGG Arg	GAT Asp	TTG	GGA G1y	GAT Asp	66C G1y	ACT	
ATC Ile	CAA	CGT Arg	AGC	TTG	GCT Ala	GAT GCT Asp Ala	TCT	
GCA Ala	TCT Ser	GAC Asp	GAG Glu	TAT Tyr	AGG		ACT Thr	
GCC Ala	TTG Leu	AAA Lys	ATG Met	GAG	CCA Pro	GAA Glu	GCG Ala	
GAG Glu	GCT	GAT Asp	GTG Val	GCA Ala	GAT Asp	CTT Leu	CAT	

i I	864	912	096	1008	1056	1116
	ω	ס	σ	10	1(
Lys	ATA Ile	AAT Asn	AAG Lys	TTT Phe	TTA	AAGGTACTTG TCATTGAGAA TACGGATTAT GGACTTGCAG AGTAATTTCC CGGAAGAGCA TATTACCACG GTTGTCCGTC AAACCCATTT AGGATACTGT
Thr	GCT Ala	ATT Ile	AAC Asn	GGA Gly	TGATTA	agta agga
Ala	GAA Glu	AGC	GCC Ala	TTC	CCA	CAG
Asn	CTT Leu	CCC	GTT Val	TCA Ser	AAG Lys	CTTG
Ile	GGT Gly	CAT	ACT Thr	AAT Asn	TTC	GGA(AAA(
AAA	GGA Gly	CTT Leu	GAC Asp	TCG Ser	GCT Ala	FTAT
ATC Ile	TCT Ser	TGG Trp	TTC Phe	ATC Ile	TCG Ser	CGGA:
GAT	GCA Ala	GGC Gly	GAG Glu	GCG Ala	TTC Phe	A TAC
AAG	GGA Gly	ACC Thr	GTG Val	GTT Val	GCT Ala	saga. ccacc
ACA	CTT	AAC Asn	TCG Ser	AAC Asn	GTG Val	ATTC
AAC ASD '	TGT	ATA Ile	CCA Pro	GTT Val	GTC Val	7G TG
AAG Lys	CAC	GGA G1y	GAG Glu	GAA Glu	TCA Ser	AGAGG
TTC 7	GGA Gly	AAG Lys	CCT	CAC His	AAC Asn	AAGGTACTTG TCATTGAGAA CGGAAGAGCA TATTACCACG
GTT 7 Val 1	ATC (Ile (ATT Ile	AAT Asn	CAA Gln	CAC His	AC ?
AAG (Lys 1	ATG /	ACT	TTC	CAG Gln	GGC G1y	CCCATTTCAC CCATGTTTGT
AAG A Lys 1	TCA Z	GCG Ala	CAA G1n	AAG Lys	GGA	CCCA

1236	1296	1348
	CCTCTGTAAA	
TCCTGTCTCC	TTTTGTTTA	AAAAAAAAA
TCCCTTTTAA	TCAAATAAGA	TCTCAAAAA
ATTATTAATT	CTTAGAAAGG	GGAAGTGCCG
AAAACTAAGG	TTTATTTTAT	GTATTGGAAA
TCTATGTAAT AAAACTAAGG ATTATTAATT TCCCTTTTAA TCCTGTCTCC AGTTTGAGCA	TGAAATTATA TTTATTTTAT CTTAGAAAGG TCAAATAAGA TTTTGTTTTA CCTCTGTAAA	ACTITIGITI GIAITGGAAA GGAAGIGCCG ICTCAAAAAA AAAAAAAAA AA

Sequence Range: 1 to 1704

40 GTG Val>		GCA Ala>		TCT Ser>	190	GAC Asp>	240	CGG Arg>	CTC Leu>		GAA Glu>
GNG Xxx		TCG Ser	140	GAC	13	ATC Ile		ATC Ile	AGG Arg		CTC
ACC Thr	06	AAT Asn		GTC Val		TTA Leu		CAG Gln	30 AGG Arg	330	GCT
30 TCC Ser		AGG Arg		GAC Asp		AGC Ser	230	GGC Gly	280 GAC AC ASP A1		AAG Lys
AGC Ser		TGC Cys	0	TCC Ser	180	ATC Ile	.,	GGC Gly	AAC Asn		AAG Lys
TGG Trp	80	GGC Gly	130	GGC Gly		$\frac{\text{GGG}}{\text{Gly}}$		TTC Phe	AAG Lys	320	$_{\rm GGG}$
20 AGC Ser		CCG		TTC Phe		AGC	220	AGG Arg	270 GGG Gly	. ,	GCC Ala
AAA Lys		CCC		GTA Val	170	GAG Glu	22	ACC Thr	GAC Asp		GTC Val
AAC Asn	7.0	GAT Asp	120	TCC	П	GGC Gly		CCC	ATC Ile	310	ATT Ile
0 GGG Gly	7	GTG Val		GTC Val		TCC Ser		TTC	260 GGA TAC Gly Tyr	m	TGC
10 AAA GGG Lys Gly		CTA		CTC	160	CTC	210	AAG Lys	2 GGA G1y		TAC Tyr
ACT Thr		GAA Glu	110	GGC Gly	16	CTC		TCC	ACG Thr		CGC Arg
CTC	09	CTA Leu	П	ATG Met		AAG Lys		GCT	250 AAC GCG Asn Ala	300	CTC
ACC Thr		GCT Ala		GGC Gly		GAA Glu	200	GAC Asp	25 AAC Asn		TGC Cys
TTA Leu		GCC Ala	0	GCC Ala	150	TAC Tyr	(1)	TTC	TTC Phe		GAT Asp
AAA Lys	50	GCG	100	CGA Arg		\mathtt{TAT}		CGC Arg	GGA Gly	90	GAC Asp

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	AGA Arg>	0	TCT Ser>	480	CCG Pro>	GCC Ala>		TGT Cys>		CGA Arg>	0.19	ATT Ile>
380	GAG Glu	430	TTC Phe		TCC	CTT Leu		GCA Ala	620	CGC Arg	6	ATC Ile
က	AAG Lys		GTC Val		AAG.ATC Lys Ile	20 CTG Leu	570	ACT Thr	Ť	ATC Ile		GCA Ala
	GAT Asp		ACC Thr	470		520 GCT CT Ala Le		TCA		CAT His		GCT Ala
0	ATT Ile	420	CTA	7.	CGG Arg	TCT Ser		ATT Ile	610	AAT	* •	GAG Glu
.370	AAG Lys		GGC Gly		CAC His	GGG Gly	260	TAT TCG Tyr Ser	9	GCC		ACT Thr
	TCC		$_{\rm GLy}^{\rm GGT}$	460	$_{\rm GLY}^{\rm GGT}$	510 ATG Met	۵,			GCT		GGA Gly
	CTC	410	ATG Met	46	AAA Lys	AAC Asn		AAC Asn		GCC Ala	650	GGA Gly
360	AGC Ser	7	GGT Gly		GAG Glu	ACA Thr	550	CCA	* 009	\mathtt{TAT}		GCT
	GAA Glu		ACT Thr		ATC Ile	500 ATT Ile	5.	GGC G1y		TTT Phe		ATT Ile
	$_{\rm GGT}$	0	${\tt GGA} \\ {\tt G1y}$	450	CTC	GCC Ala		ATG Met		TGC	640	ATG Met
350	GGC Gly	400	GTT Val		AAT Asn	TAT Tyr		CTG	590	TAC	9	CTC
m	CTC		CTA		CAG Gln	490 T CCC e Pro	540	GGT Gly	٠,٠	AAC Asn		GAC
	GAT Asp		GTG Val	440	GTT Val	49 ATT Ile		TTG		TCC		GCT
0	TCC	390	$_{\rm GGA}$	7	666 Gly	TTC		GAT Asp	80	ACT Thr	630	GAG Glu
340	AAT Asn		GCT Ala		GAC Asp	TTT Phe	30	ATC Ile	25	GCT		GGC Gly
									•			

SUBSTITUTE SHEET (RULE 26)

FIGURE 2 3/5

720	AGG Arg>	GAT Asp>		TTG Leu>		GGA Gly>	910	GAT Asp>	096 *	GGG Gly>	ACT Thr>
	CAA Gln	CGT Arg		AGC Ser	860	TTG Leu	6	GCT Ala		GCT Ala	TCC Ser
	TCT Ser	50 GAC ASP	810	GAG Glu	ω	TAT Tyr		AGG Arg		GAT Asp	1000 GCG ACT Ala Thr
710	TTA Leu	760 AAG G2 Lys A8		ATG Met		GAA Glu		CCA Pro	950	GAA	100 GCG Ala
7	GCT	GAT Asp		GTT Val	850	GCA Ala	900	GAT	0,	CTG	CAT His
	AGG Arg	TGG Trp	800	TTG Leu	8	ATT Ile		ACT Thr		AGT Ser	GCT
0	TGC Cys	750 CCG Pro	ω	GTA Val		ATT Ile		ATG Met	940	AGC	990 AAT Asn
700	GCC Ala	AGG Arg		GGA Gly		CCG Pro	890	CAT His	õ	GAG Glu	ATA Ile
	GTT Val	TCA Ser	0	GCT Ala	840	GCG Ala	w	TAT Tyr		ATT Ile	TAC
	TTC Phe	740 GCC	790	$_{\rm GGG}$		GGA Gly		GCT		TGC	980 GTC AAT Val Asn
069	GGA Gly	7 ACT Thr		GAA Glu		CGA Arg	880	GAT Asp	930	TCT	
	GGA Gly	CAG Gln		GGC Gly	830	AAA Lys	88	TGT Cys		TCC	GAG Glu
	rta Leu	730 GAC CCT ASP Pro	780	ATG Met	ω	ATG Met		AAT Asn		GTC Val	970 CCT GAA Pro Glu
089	GGG	73 GAC ASP		GTG Val		GCA Ala		GTC Val	920	$_{\rm G1y}^{\rm GGT}$	9. CCT Pro
9	ATT Ile	GAT Asp		TTT Phe	820	CAT	870	GCA Ala	0.	CTT Leu	TCA
	CCA Pro	AAT Asn	70	GGT Gly	8	GAA Glu		$_{\rm GGT}$		GGG G1y	GTC Val

FIGURE 2

	r TTC AAG l Phe Lys>	1100	c GGA CAC e Gly His>	1150	T AAG GGA e Lys Gly>	1200	T CCC GAG n Pro Glu>	A CAT GAA n His Glu>		C AAC TCA s Asn Ser>	1340	A AAT GCA
1050	AAG GTT Lys Val		ATG ATC Met Ile		ACA ATT Thr Ile	1190	TTC AAT Phe Asn	1240 CAG CAA Gln Gln	1290	GGC CAC Gly His		GGT TCA AAT
	AAG Lys	1090	AAG TCG Lys Ser	1140	GCG Ala	11	CAA Gln	AAG Lys		GGA Gly	1330	TTA CTC
1040	ATC AIL	10			C ATT a Ile		A AAC e Asn	0 C AAG n Lys	1280	A TTC y Phe	Ħ	TGA TT
7	AAT GCC Asn Ala		GCA ACT Ala Thr	0	GAA GCC Glu Ala	1180	AGC ATA Ser Ile	1230 GCC AAC Ala Asn		TTC GGA Phe Gly		CCA TG Pro
0	ATA AA Ile As	1080	AAT GC Asn A]	1130	CTT G	` '	CCC A(GTT G Val A	. 0	TCA	1320	AAG Lys
1030	GAG Glu	7	ATC Ile		$_{\rm GGT}$		CAT	20 ACA Thr	1270	AAT	T	TTC Phe
	GCC		ACA Thr	1120	4 GGG	1170	CTT Leu			C TCA e Ser		A GCC r Ala
O *	GAT CTT Asp Leu	1070	A ATC u Ile	11	A TCA a Ser		C TGG Y Trp	A TTC u Phe	0 +	T ATC a Ile	1310	TTC TCA Phe Ser
1020	G GAT y Asp		G GAA s Glu		A GCA Y Ala	_	ACC GGC Thr Gly	1210 GTG GAA Val Glu	1260	GTT GCT Val Ala		GCT TTC Ala Phe
	T GGG a Gly		C AAG Ir Lys	0.	CTT GGA Leu Gly	1160	ACC AC Thr Th	1 TCA GT Ser Va		AAT GTT Asn Val		GTA GC Val A
	CTT GCT Leu Ala	1060	AAC ACC Asn Thr	1110	TGT CTT Cys Leu		ATA AC Ile Th	CCA TC Pro Se	0	GTG AA Val As	1300	GTT G
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SUBSTITUTE SHEET (RULE 26)

FIGURE 2 5/5

CCCTTGTCAA TGGCATTTAA GATAAGCTTA TAAAAAAAA AAAAAAAA AAAACTCGAG TATTAGAAAG AACGAGGCAA GATTTTTGTTT CATGTTTGTG TTTGTATTAC TTTCTTTTTG GGGGGGCCCG GTACCCAATT CGCCCTATAG TGAGTCGTAT GACAATTCAC TGTCCGTCGG AATTIGIIGC IGAGACAGIG AGCIICAACI IGCAGAGCAA ITITITACAI GCCIIGICGI CGGAAGAGCG TAATACCGGG ATAGTTCCTT GATAGTTCAT TTAGGATGTT TTACTGCAAT AATCGAAGAT TATTTCCATT CTAATCCAGT CTCCGNCGAG TTTGAGAATC TATCTGTTTG

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* 09	CCGCTCTAGA ACTAGTGGAT	120	GCTCAGGTGT	ACG TGG	·	CGT TCC Arg Ser	09	CTC TCC Leu Ser	310	CCT TGC Pro Cys	360	TTC GGA Phe Gly
20	CTCTAGA A	110	GGTCGGCTCA G	160 T TTC TGT o Phe Cys	210	GAC CCA Asp Pro	260	AGG ACT Arg Thr		CTC GAT Leu Asp	350	TCC CTC Ser Leu
0		0		TCC CCT Ser Pro		C AAC p Asn	0	C CGG g Arg	300	A TGC n Cys	m	C GCT e Ala
40	GCGGTGGCGG	100	TTCTTACTTG	50 GCG Ala	200	TCC GAC Ser Asp	250	CGT CGC Arg Arg		TTC CAA Phe Gln		GGA TTC Gly Phe
0	505 C	0		l G GTT t Val		TCA		TCC Ser	290	ACC Thr	340	AAC Asn
30	GAGCTCCACC	06	GGCACGAGTT	140 TCT TGC ATG Ser Cys Met	190	CCC ACT Pro Thr	240	CGC CTC Arg Leu	2	GGA TCC Gly Ser		GGG GAT Gly Asp
20	CTG GA	80		14 GCT TCT Ala Ser	•	TGC ATG Cys Met		3G CTC	280	CTC CGC Leu Arg	330	TTC CTC Phe Leu
	ACAAAAGCTG		GCAGGAATTC	ACC Thr	180	GCA Ala	230	AAG CGG Lys Arg	28	TCC		CGC
10	actaaaggga <i>i</i>	70	LOGOGOCI	130 ATG GCG Met Ala	(-)	GTA GCT Val Ala		TCC CAC Ser His	0,	CAT TGC His Cys	320	CAG CAA Gln Gln
	ACTAA		ລລລລ	TCCA	170	CTC G	220	CTT I	270	TCC C		AAC C Asn G

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ACT Thr		GAA Glu		GTG Val		TAC Tyr	* 009	AAC Asn	TCT Ser	•	GAC Asp
CGC		CAG Gln	0	GTT Val	550	GTT Val	Q	GAG Glu	AAG Lys		ATG Met
GGC G1y	450	GCA Ala	200	GTA Val		GAT Asp		ATA Ile	ATC	069	AGG Arg
400 CTC Leu	4	CCT Pro		CGA Arg		CCC Pro	290	GAG Glu	640 GAG Glu		GAG Glu
AGG Arg		CAA Gln		caa agg gln arg	540	GAC	5.	AGT Ser	GGA G1y		TCC
CTG	0	ATG Met	490		.,	CAT		ATA Ile	GCC	089	TTC
390 CAC His	440	GCT Ala		AAG Lys		GGC Gly		GGC Gly	630 ATT Ile	9	AAG Lys
3 GGC G1y		GTG Val		ACC Thr	530	CTA	580	AGT	AGA Arg		CCA
CGC Arg		GCT Ala	480	CCT GCT Pro Ala	5.	CCT Pro		ATA Ile	ACG		GCC Ala
0 AAT Asn	430	ATG Met	7			ACT Thr		GGA G1y	20 CCC Pro	670	GTG Val
38 TCA Ser		GTC Val		AAA Lys		GTG GTG Val Val	570	CTA GAC (Leu Asp (620 TTT CC Phe Pi		TGG Trp
CGT Arg		GAG Glu	470	AAG Lys	520	GTG Val	2,	CTA	CAG Gln		GGC Gly
CTT Leu	420	GGG Gly	47	AAT Asn		GGC Gly		CTC	TCT Ser	099	GAT Asp
370 CCT Pro	Þ	TCC Ser		ACA Thr		ATG Met	09	AAT Asn	610 TGC Cys		ACA Thr
AAG Lys		CAT His		TCC	510	GGT Gly	2	AAC Asn	GAC Asp		TCC
TCC	410	TCC	460	GTC Val	· ·	ACA Thr		TAC	TTC Phe	650	TTT

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GAT Asp		TGT Cys	840 *	GAT Asp	TGT Cys		GAC Asp		ACA Thr	:	GAA Glu
GCA	190	AAG Lys	8	AGC Ser	TTT Phe		ATG Met	086	TGT GCA Cys Ala	1030	GGC Gly
TTA Leu		AGA Arg		TTC Phe	CCC	9.30	GCA Ala	9			AAA Lys
GCA Ala		AAA Lys	830	GTA	880 AGT Ser		CTT Leu		GCC Ala		ATC Ile
AAA Lys	780	AAT Asn	8	AAG Lys	ATC Ile		ATT Ile		ACT Thr	1020	AAC CAC ATA Asn His Ile
AAG Lys	1	CTC		ATG Met	AAG Lys	920	TCC GCT Ser Ala	970	TCA	. 4	CAC
GGC Gly		GAG Glu		$_{\rm GGT}$	870 TAT AAG Tyr Lys	6			ATA Ile		AAC
GCA Ala	0	AAA Lys	820	GGC Gly			GGA Gly		TCG	1010	GCG
ACT Thr	770	ATG Met		TTG	TCA		ATG Met	096	\mathtt{TAT}	10	GCT
CTG		GCG		GGA Gly	860 AGG ACT Arg Thr	910	AAT Asn		AAC Asn		AAT
* ATG Met		GAT Asp	810	TCC			ACA Thr		CCT		CTG
TAC	760	GAA Glu	ω	GGC	CTG		ACC Thr	950	GGC Gly	1000	ATA Ile
CTT		ACT Thr		ATT Ile	GCT	006	TCT	9	ATG Met		TGT
ATG Met		ATC Ile	800	CTC	850 GAA G1u	-	TTT Phe		TGG Trp		TTC
TTC Phe	750	GGA Gly	8(GTT Val	ATT Ile		CCT		GGA Gly	066	AAC
AAG Lys	-	${\tt GGT} \\ {\tt G1y}$		GGA Gly	TCC	890	GTA Val	940	${ m TTG}$		AGT

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1080	GTT Val	AAT Asn		TTT Phe		CAT His		AGT Ser	1320	GCT Ala	TCG
10	CCT	AAT Asn		GGA Gly	0	GAG Glu	1270	$_{\rm GGG}$	=======================================	GGA Gly	GTC Val
	TTA Leu	AGG Arg	1170	GAT Asp	122	TTA Leu	H	$_{\rm GLY}^{\rm GGT}$		GAA Glu	GGA G1y
0	GTT Val		11	CGT Arg		GAG Glu		CTA Leu	01	CCT	1360 ; TCC
1070	GCC Ala	1120 TCA CAG Ser Gln		AAT Asn		GAG Glu	1260	TTT Phe	1310	CAC His	CAG
	GCG Ala	TTG Leu	0.0	AGT Ser	1210	CTT	13	GAA Glu		CCT	GCT Ala
	GAT Asp	1110 3A GCT 7g Ala	1160	GAC	•	CTT		GCG Ala		GAG Glu	350 TTG Leu
1060	TCG Ser	11 CGA Arg	. •	TGG Trp		TTA Leu	20	\mathtt{TAT}	1300	ACC	1350 GCC TTC Ala Leu
1	66C Gly	TGC Cys		CCA Pro	1200	GTT Val	125(ATT Ile		ATG Met	AAG Lys
	GGT Gly	1100 GTA GCA Val Ala	1150	AGA Arg	H	GGA Gly		ACC Thr		CAC His	40 GAG Glu
1050	TGT Cys	1100 GTA G Val A	(1	TCG Ser		GCT Ala		GCA	1290	TAC Tyr	1340 ATA GA Ile G
10	CTT Leu	TTC		GCT Ala	06	GAA GGA Glu Gly	1240	GGT Gly	H	GCC Ala	TGC Cys
	ATG Met	$_{\rm GGT}$	1140	AAA Lys	119(GAA Glu	, ,	AGA Arg		GAC Asp	CTC
0	* ATG Met	1090 GGA Gly	Ξ	ACC Thr		GGA Gly		AAA Lys	30	TGC Cys	1330 ATC Ile
1040	GAC Asp	1 TTG Leu		CCT Pro		ATG Met	1230	AAG Lys	1280	ACT Thr	GTG Val
	GCA Ala	GGT G1y	1130	GAC Asp	1180	GTG Val	12	GCA		TTC Phe	GGT Gly

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	GCT Ala		AAC Asn		CTT	1560	AGG Arg	GGC		GTC Val		TCC
		0	CAA Gln	1510	CTT	72	ATA Ile	GAA Glu	•	AAG Lys	00	TCA
	ACT Thr	1460	GGC Gly	7	CAC		GCA Ala	GAC Asp	1650	CTG	1700	AAC
	TCC		TTC Phe		GGT Gly	1550	CAG Gln	1600 GAA GAC CCG Glu Asp Pro	H	AAA Lys	•	CAT
	ACT Thr		TGT Cys	1500	ATC Ile	15!	GTT Val	1 GAC ASP		GAG		GGC Gly
	GCA	1450	CAC	4	ATG Met		GTA Val		40	AAG Lys	1690	GGC Gly
	CAT His	П	GCC Ala		TCG Ser		GCA	1590 AAT TTG Asn Leu	1640	AAG Lys		TTC
	GCG Ala		CTC Leu	06	AAA Lys	1540	GTT Val			CCT Pro		TTT GGG Phe Gly
	AAT Asn	1440	GCT	1490	ACC Thr		GCA Ala	ATT Ile		GGC Gly	1680	TTT
	ATA Ile	14	CAA Gln		TCC		GAA Glu	80 AAT Asn	1630	GTC Val	⊶.	TCA
	TAC Tyr		TAC Tyr		AAT	1530	GTA Val	1580 CCA AA Pro As		CTC		AAT Asn
	AAT Asn	0	GAA Glu	1480	GTG Val	Ħ	GGC Gly	CAT		CTG	1670	TCC Ser
*	GTA	1430	AAG Lys	•	AGA Arg		$_{\rm GGT}$	ATC Ile	1620	AAA Lys	16	TTG
	GAC Asp		ATC Ile		CTG	0	GGA GCT Gly Ala	1570 TGG Trp	Ţ	GCA Ala		GGT Gly
	GAA Glu		GAT Asp	1470	GAG Glu	1520	GGA Gly	1 GGA Gly		GAT Asp		GTC
	AGG Arg	1420	GGA Gly	14	AGT Ser		GGA Gly	ACA Thr	610	GTG Val	1660	AAG Lys
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	AAAAA	AAACCACATC TCAGTATGCA AAATAAAAA AAAAAAAAA AAAAAA	AAATAAAAA	TCAGTATGCA	AAACCACATC
		2040	2030	2020	2010
ATATTTTGAA	TTTGTAATGC	ACATGTTCGT TATCGGATCA ATGTGTTTCT TCTAAGATCA TTTGTAATGC ATATTTTGAA	ATGTGTTTCT	TATCGGATCA	ACATGTTCGT
2000	1990	1980	1970	1960	1950
CTTTTCGAAT	TATTTCGAG	TATITICITC TICITITGAG AGCITITAACC GAGGTAGICG TATITICGAG	AGCTTTAACC	TTCTTTTGAG	TATTTTCTTC
1940	1930	1920	1910	1900	1890
TTGTCCCTTT	AGATCACTGC	GGGGATGCCA AAGATACTCC TTGCCGGTAT TGGTGTTAAG AGATCACTGC TTGTCCCTTT	TTGCCGGTAT	AAGATACTCC	GGGGATGCCA
1880	1870	1860	1850	1840	1830
GGCTACTCGA	GAGATAGACC	GAACTCATGC ACGTTAGTAG CTTCTTATGC CTCTGAAACC GAGATAGACC GGCTACTCGA	CTTCTTATGC	ACGTTAGTAG	GAACTCATGC
1820	1810	1800	1790	1780	1770
TAG A AAAGAGTCTG TGGAAGCCGA GAGTCTTTGA ***	TGGAAGCCGA	. AAAGAGTCTG	AAC Asn	r GCC CCC TGC Ala Pro Cys	ATA CTA TTT Ile Leu Phe
- 1760	1750	1740	1730	1720	1710

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	09	CTACACCTCC	120	GCTCAATCGA	180	CTGCACAGGA AGTTACCACA	ATG Met>		AAT Asn>		TGT Cys>	370	ACA Thr>
.*		TACA		CTCA		GTTA	GGA Gly		AAT Asn	320	GAT Asp	ς.	TCC
	20		110		170	GA A	220 GTG ACT Val Thr	270	TAC	(.,	TTT Phe		TTC
		TGAC	Н	ອອວລາ	П	ACAG	2 GTG Val		TTC		ACC Thr		AAG TCT Lys Ser
		GCCATGACTA		GGCACCGGAG		CTGC	GTT Val		GTT Val	310	GAG	360	
	40		100	GCA	160	AAC	GTA Val	260	GAT	·κή	ATA Ile		ÀTC Ile
		TTCGAGCCCT		ACCC		GCTCTGCAAC	210 CGA (CCT Pro		GAG Glu		GAG Glu
		TTC	_	; ACC			CGG Arg		GAC	٠	AGT	350	GCT GGA Ala Gly
	30	CCTCGCCTGC	90	TCGGATCCAG GCCCATCCGC ACCACCCGCA	150	AATGGCTGTG	200 ATC AAA CAG Ile Lys Gln	250	CAT His	300	ATA Ile		GCT Ala
		TCGC		CCCA		4TGG(200 AAA Lys	2	GGC Gly		GGC		AGA ATT Arg Ile
	20	ra cc	80	4G G(140	3C A			CTA Leu		AGT	340	
	(7	TCT	ω	ATCC2	1,	3GAG(AGT		CCT. Pro	290	ACG Thr	3	CCT ACG Pro Thr
-		CGGCACGAGG TCACCTCTTA		rcgG2		CCGGGGGAGGC	190 AAG CCA AGT Lys Pro Ser	240	A T	,	GGA Gly		
1921	10	YGG · T	7.0	rgT 1	130	CTT (GTG Val		GAT Asp		TTT Phe
1 to		ACG		SCATCCTTGT	(-1	GCTTCCCCTT	AAG Lys		GTG Val	280	CTT Leu	330	CAA Gln
ıge:		0990		GCAT		GCT	AAG Lys	230	GGT Gly	2	CTG		GCT Ala
Rang													
Sequence													
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FIGURE 4

	420	ATG Met>	ATC Ile>		CTC Leu>		GAA Glu>	0.	TTC Phe>	* 099	TGG Trp>	rrr [.] Phe>
,	,	TTC Phe	GGA		GTT Val	260	ATT Ile	610	CCT Pro		GGA Gly	AAC Asn
		AAG '		510	GGA Gly	ഹ	GCC Ala		GTA Val		TTG Leu	700 G AGT r Ser
	410	GAC A	460 AAT GGT Asn Gly		TGC Cys		GAT Asp		TGT Cys	029	GAC	70 ACG Thr
	4	ATG (ACA		AAA Lys	20	AAT Asn	009	TTT Phe	Ψ	ATG Met	GCA Ala
		AGG A	TTA	200	AGA Arg	52	TTC Phe		CCC Pro		GCA Ala	TGT Cys
	0	AAG	450 GCA Ala	5	AAA AGA Lys Arg		GTA Val		AAT Asn	640	CTT	690 GCT Ala
	400	TCC	AAA Lys		GAT Asp		AAG Lys	290	ATG Met	9	ATG	ACT Thr
		CTC	AAG Lys	0	CTA	540	ATG	Lr)	AAG Lys		GCT	TCT
		AAG Lys	440 GGC Gly	490	GAG Glu		GGA Gly		AAG Lys		TCA	680 ATA Ile
	390	CCG	4 GCC Ala		AAA Lys		$_{\rm GLy}^{\rm GGT}$	280	\mathtt{TAT}	630	GGA Gly	TCG
		GCC Ala	ACT Thr		ATG Met	530	ATG Met	58	TCA Ser		ATG Met	TAC Tyr
		GTG Val	rg G	480	GTG Val	ΓΩ	GCA Ala		ATT Ile		AAT Asn	670 CCC AAC Pro Asn
	380	TGG	430 ATG C1 Met Le		GAT Asp		TCA Ser		AGG Arg	620	ACA Thr	
	М	$_{\rm GGT}^{\rm GGT}$	TAC		GAA Glu	0	66C Gly	570	CTA	v	ACC Thr	GGC
		GAT Asp	CTT	470	ACC Thr	520	ATT 11e		GCC Ala		GCT Ala	ATG Met

FIGURE 4 2/6

	GTG Val>		GGA Gly>	0	ACT Thr>	006	GGG Gly>	AAA Lys>		TGC Cys>	•	ATT Ile>
	GAT ASP	800	ATG Met	850	CCT		ATG Met	AAG Lys		ACT Thr	1040	GTG Val
750	GCA	∞	GGT Gly		GAC		GTT Val	940 CAT GCA His Ala	066	TTC	1(GGA Gly
	GAA		ATT Ile		GCC	890	TTT Phe			AGT		GCT Ala
	GGC Gly	0.	CCT Pro.	840	AAT Asn	ω	GGA Gly	GAG		GGA Gly	30	GGA Gly
740	AGA Arg	790	ATA Ile		AGA Arg		GAT Asp	TTA Leu	086	${\tt GGT}\\ {\tt G1y}$	1030	GAT
7	ATC Ile		ATC Ile		CAG Gln	880	CGT	930 GAG Glu		CTA		CCT
	ATA Ile		GTA Val	830	TCA	88	AAT Asn	GAG		TTT Phe		CAC His
. 0	CAC	780	GCG Ala	ω	TTG		AGT Ser	CTA	016	GAA Glu	1020	CCT
730	AAC Asn		GAT Asp		GCT		GAC Asp	920 CTA CTA Leu Leu	9	GCA	,	GAG Glu
	GCG Ala		TCA	0	CGA Arg	870	TGG Trp			TAC		ACC Thr
	GCT Ala	770	GGC Gly	820	TGC		CCA	GTG Val		ATT Ile	1010	CAC ATG His Met
720	AAT Asn	. 7	666 Gly		GCA Ala		AGA Arg	10 GGA G1y	096	ACT Thr	H	CAC
	CTG Leu		TGC		GTT Val	098	TCA	910 GCT GGA Ala Gly		GCG Ala	٠.	TAC
	ATC Ile	0	CTT Leu	810	TTT Phe	ω	GCT Ala	GGA G1y		GGT Gly	00	GCC
710	TGT	160	ATG Met		GGT Gly		AAA Lys	GAA Glu	950	AGA Arg	1000	GAT Asp
								•				

FIGURE 4 3/6

0	GAC Asp>	1140	ATC Ile>	TTA Leu>		GCC Ala>		TGG Trp>	30	ACC Thr>	1380	GTC GGT Val Gly>
1090	GAA Glu	Ε,	GAT Asp	GAG Glu		GCA Ala	1280	$_{\rm GGG}$	1330	GAT	` '	GTC Val
	AGG Arg		GGA Gly	1180 AAC AAC Asn Asn	1230	GGA Gly	13	ACT Thr		GTG Val		AAG Lys
	TCT	1130	GCT Ala	1180 AAC AJ ASD A	П	CTC		AGG Arg		66C 61y	1370	AAC ATT Asn Ile
1080	GTC Val	11	CCA Pro	CAA Gln		CTT Leu	0.0	GCA ATA Ala Ile	1320	GAA Glu	ä	AAC Asn
-	GGA Gly		ACT Thr	66C 61y	1220	CAC His	1270	GCA Ala		GAT Asp		CTG
	TCA	0.	ACA TCC Thr Ser	1170 TGT TTC Cys Phe	12	$_{\rm GLY}^{\rm GGT}$		CAG Gln		CCA	0.5	AGA Arg
1070	CAG Gln	1120	ACA Thr			ATT Ile		TCA GTA GTT Ser Val Val	1310	GAA AAC Glu Asn	1360	GAG
10	GCT		GCC	CAC His	0	ATG Met	1260	GTA Val	H			AAG Lys
	TTG Leu		CAT His	1160 T ATC	1210	TCA	•			TTG		AAG Lys
0	GCT Ala	1110	GCA Ala	<u>5</u> 3		AAA Lys		GTT Val	00	AAT Asn	1350	CCT
1060	AAG Lys	П	AAT Asn	GCT Ala		ACC Thr	1250	GCA Ala	1300	ATT Ile	\- 1	GGC G1y
	GAG Glu		ATA Ile	CAA Gln	1200	TCT Ser	13	GAA Glu		AAT Asn		GTG Val
	ATA Ile	1100	TAC Tyr	1150 TAC CAA Tyr Gln	1	AAT Asn		GTG Val		CCG	1340	CTC
1050	TGC	11	AAT Asn	GAG Glu		GTG Val	0	GGT GGT Gly Gly	1290	CAT His	Ħ	TTG
	CTC		GTA Val	AAA Lys	1190	AAA Lys	1240	GGT Gly	П	ATC Ile		AAA Lys

FIGURE 4

ATA CTC TTC Ile Leu Phe>	1480	TCAAA	1540	CATGCCCATG	1600	GGCGACACAG	1660	TTTCTGAAAT	1720	AGTCAGTGAA GAAGAGAACA	1780	TTTATCGCCG	1840	ATCATTGGAG
1420 TCG TCC Ser Ser	1470	TAG GGCGTTT CATGTGGA ATTCTACTCA ATCTATCAAA ***>	1530	TGAGGACTCC AGCATGTTGG TAGCTCCTTA CGTCTCTAGA CATGCCCATG	1590	GAGTACTCAT	1650	CTATTCATTA TCCCATTTTT TTTCTGAAAT	1710		1770	TGCTCTCTAT	1830	TTTTGTGGGT TAAAATTTGT AAAACTAGAC GACTGGTTTG TTTTCTCTTG ATCATTGGAG
1410 GGG CAC AAC Gly His Asn	1460	GTGGA ATTCT	1520	TAGCTCCTTA	1580	ATGACGGATT	1640	CTATTCATTA	1700	CGTTTCATCG	1760	CCCTTTGTTT	1820	GACTGGTTTG
1400 GGG TTT GGT G Gly Phe Gly G	1450	CGTTT CATGT	1510	AGCATGTTGG	1570	CGGGAGCTGT AGTCGGAACC	1630	TGTTAGAGCA	. 1690	CTCCCTCCTT ACGGTAGTTG TACTTTCGAG CGTTTCATCG	1750	TAACCATTTG	1810	AAAACTAGAC
TCA TTC Ser Phe	1440	AAC Asn	1500	TGAGGACTCC	1560	CGGGAGCTGT	1620	TTGCTAGAAT	1680	ACGGTAGTTG	1740	GGGCACGTAG	1800	TAAAATTTGT
1390 TTG TCT AAT Leu Ser Asn	1430	GCC CCT TAC Ala Pro Tyr	1490	GCTGAAGTTT	1550	AGTTTTGTGT	1610	GATATACTCC	1670	CTCCCTCCTT	1730	AAGCTAACTC	1790	TTTTGTGGGT

FIGURE 4 5/6

FIGURE 4 6/6

ATGTATGGCC ATATTTGCCT TTCATTGATG ATAAAAAAA AAAAAAAA AAAAAAAAA 1890 1880 1870 1860 1920 1910

AAAAAAAA AAAAAAAA A

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09	120	169	217	265	313	361	409	457	505
CTGGTACGCC TGCAGGTACC GGTCCGGAAT TCCCGGGTCG ACCCACGCGT CCGTCTTCCC	ACTCCGATCG TTCTTCTTCC ACCGCATCTC TTCTCTTCTC	CGCCGCC ATG CAT TCC CTC CAG TCA CCC TCC CTT CGG GCC TCC CCG CTC Met His Ser Leu Gln Ser Pro Ser Leu Arg Ala Ser Pro Leu 1	GAC CCC TTC CGC CCC AAA TCA TCC ACC GTC CGC CCC CTC CAC CGA GCA Asp Pro Phe Arg Pro Lys Ser Ser Thr Val Arg Pro Leu His Arg Ala 15	TCA ATT CCC AAC GTC CGG GCC GCT TCC CCC ACC GTC TCC GCT CCC AAG Ser Ile Pro Asn Val Arg Ala Ala Ser Pro Thr Val Ser Ala Pro Lys 35	CGC GAG ACC GAC CCC AAG AAG CGC GTC GTG ATC ACC GGA ATG GGC CTT Arg Glu Thr Asp Pro Lys Lys Arg Val Val Ile Thr Gly Met Gly Leu 55	GTC TCC GTT TTC GGC TCC GAC GTC GAT GCG TAC TAC GAC AAG CTC CTG Val Ser Val Phe Gly Ser Asp Val Asp Ala Tyr Tyr Asp Lys Leu Leu 65	TCA GGC GAG AGC GGG ATC GGC CCA ATC GAC CGC TTC GAC GCC TCC AAG Ser Gly Glu Ser Gly Ile Gly Pro Ile Asp Arg Phe Asp Ala Ser Lys 80	TTC CCC ACC AGG TTC GGC GGC CAG ATT CGT GGC TTC AAC TCC ATG GGA Phe Pro Thr Arg Phe Gly Gly Gln Ile Arg Gly Phe Asn Ser Met Gly 95	TAC ATT GAC GGC AAA AAC GAC AGG CGG CTT GAT GAT TGC CTT CGC TAC Tyr Ile Asp Gly Lys Asn Asp Arg Arg Leu Asp Asp Cys Leu Arg Tyr 115

553	601	649	697	745	793	841	888
GCC Ala	GGG Gly	CTT Leu	GCC Ala 190	ATG Met	TGC Cys	ATG Met	GGC Gly
GGT Gly	GTT Val	TCT Ser	TAT Tyr	CTG Leu 205	TAC	CTT Leu	GGA Gly
CTC Leu 140	CTG	CAA Gln	CCC Pro	GGT G1y	AAC Asn 220	GAT Asp	TTG
GAT Asp	GTG Val 155	GTT Val	ATC Ile	CTC	TCC Ser	GCT Ala 235	666 G1y
GCC Ala	GGA Gly	GGG Gly 170	TTC Phe	GAA Glu	ACT Thr	GAG Glu	ATT Ile 250
GAC Asp	GCC Ala	GAC Asp	TTC Phe 185	ATT Ile	GCC Ala	$_{\rm G1y}$	CCA Pro
GAG Glu	AGA Arg	TCT	CCT Pro	GCT Ala 200	TGT Cys	CGT Arg	ATT Ile
CTT Leu 135	GAG Glu	TTC Phe	ACC Thr	CTC	GCA Ala 215	CGC Arg	ATC Ile
TCT Ser	AAG Lys 150	GTC Val	ATC Ile	CTG	ACT Thr	ATC 11e 230	GCA Ala
AAG Lys	GAC Asp	ACT Thr 165	AAA Lys	GCC Ala	TCC Ser	CAT His	GCC Ala 245
AAG Lys	ATC Ile	CTG	CGG Arg 180	TCT Ser	ATT Ile	AAT Asn	GAG Glu
666	AAG Lys	GGT Gly	CAC	GGG Gly 195	TCA Ser	GCT Ala	ACT Thr
GCC Ala 130	TCC Ser	GGT Gly	GGT Gly	ATG Met	TAT Tyr 210	GCT Ala	GGC
GTC Val	CTC Leu 145	ATG Met	AAG Lys	AAC Asn	AAC Asn	GCT Ala 225	GGA Gly
ATT Ile	CGC Arg	GGA G1y 160	GAG Glu	ACA Thr	CCA Pro	CAT His	GCT Ala 240
TGC Cys	GAC	ACA Thr	ATC Ile 175	ATT Ile	GGC G1y	TTC Phe	ATT Ile

FIGURE 5 2/4

937	586	1033	1081	29	.76	1224	1272
6	σ	10	10	112	117	12	
ACT Thr 270	GAA Glu	CGA	GAT Asp	TCT Ser	GTC Val 350	GCC Ala	AAA Lys
CAG Gln	GGT G1y 285	AAA Lys	TGT Cys	TCC	GAG Glu	CTC Leu 365	ATC Ile
CCT Pro	ATG Met	ATG Met 300	AAC Asn	GTC Val	GAA Glu	GAT Asp	GAT Asp 380
GAC Asp	GTG Val	GCA Ala	ATC Ile 315	GGT.	CCT Pro	666 Gly	AAG Lys
GAT Asp	TTT	CAT His	GCA Ala	CTC Leu 330	TCA Ser	GCT Ala	ACA Thr
AAC Asn 265	GGT Gly	GAA Glu	GGT Gly	GGT Gly	GTC Val 345	CTA Leu	AAC
AGG Arg	GAT Asp 280	TTG Leu	GGA Gly	GAT Asp	66C Gly	ACT Thr 360	AAG Lys
CAA Gln	CGT Arg	AGC Ser 295	TTG Leu	GCT Ala	GCT Ala	TCT Ser	TTC Phe 375
TCT Ser	GAC Asp	GAG Glu	TAT Tyr 310	AGG Arg	GAT Asp	ACT Thr	GTT Val
CTG	AAA Lys	CTG	GAG Glu	CCA Pro 325	GAA Glu	GCG Ala	AAG Lys
GCT Ala 260	GAT Asp	GTG Val	GCA Ala	GAC Asp	CTT Leu 340	CAT His	AAG Lys
AGG Arg	TGG Trp 275	TTG Leu	ATT Ile	ACT Thr	AGC Ser	GCT Ala 355	ATC Ile
TGC Cys	CCC	GTG Val 290	ATT Ile	ATG Met	AGT Ser	AAT Asn	GCC Ala 370
GCT Ala	AGG	GGA Gly	CCT Pro 305	CAC	GAG Glu	ATA Ile	AAT
GTG Val	TCT	GCT Ala	GCA Ala	TAT Tyr 320	ATT Ile	TAC Tyr	ATA Ile
TTT Phe 255	GCC	GGT Gly	GGA Gly	GCT Ala	TGC Cys 335	AAT Asn	GAG Glu
					•		

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1320	1368	1416	1464	1512	1569	1629	1689	1712
ATT AAT GCA ACT AAG TCA ATG ATC GGA CAC TGT CTT GGA GCC TCT GGA Ile Asn Ala Thr Lys Ser Met Ile Gly His Cys Leu Gly Ala Ser Gly 385	GGT CTT GAA GCT ATA GCG ACT ATT AAG GGA ATA AAC ACC GGC TGG CTT Gly Leu Glu Ala Ile Ala Thr Ile Lys Gly Ile Asn Thr Gly Trp Leu 400	CAT CCC AGC ATT AAT CAA TTC AAT CCT GAG CCA TCC GTG GAG TTC GAC His Pro Ser Ile Asn Gln Phe Asn Pro Glu Pro Ser Val Glu Phe Asp 415	ACT GTT GCC AAC AAG AAG CAG CAA CAC GAA GTT AAT GTT GCG ATC TCG Thr Val Ala Asn Lys Lys Gln Gln His Glu Val Asn Val Ala Ile Ser 435	AAT TCA TTT GGA TTC GGA GGC CAC AAC TCA GTC GTG GCT TTC TCG GCT Asn Ser Phe Gly Phe Gly Gly His Asn Ser Val Val Ala Phe Ser Ala 450	TTC AAG CCA TGA TTACC CATTTCACAA GGCACTTGTC ATTGAGAGTA CGGTTGTTCG Phe Lys Pro 465	TCAAACCCAT TTAGGATACT GTTCTATGTA AAAAAAGTA AGGATTATCA CTTTCCCTTC	TAATCCTGTC TCCAGTTTGA GAATGAAATT ATATTTTTT TAAAAAAAA AAAAAAGGGC	GGCCGCTCTA GAGGATCCAA GCT

IGURE 5

Sequence Range: 1 to 1802

0 9	TTATCTCCGC	110 TCC CCT TCC Ser Pro Ser		TCC	210	CGT		CGG Arg		GTC Val	CTA Leu
	'TAT'	11(TCC (Ser]	160	TCC Ser		ATC Ile		AAG Lys	,	GAC	350 ATC AGC Ile Ser
20		CAC 1 His 9		CCC Pro		GTC Val		AAG Lys	300	GGC TCC Gly Ser	35 ATC Ile
	SCCT	CTC C Leu H		TCC Ser	0	CCC	250	CCC Pro	(*)	GGC GLy	GGC
	rctt(100 TCC C	150	AAT Asn	200	CTC		GAC Asp		TTC Phe	AGC
40	ATT	1 CAA T Gln S		CTC		AGC Ser		TĊĊ Ser	0	GTC Val	340 GAG Glu
	rttC2			CGC		GCC	240	GAG	290	TCC	GGC Gly
	ACA!	C A A		TTC	190	CGC		CGC		GTC Val	TCC
30	SACC	2522 06	140	CCC		CGT		AAG Lys		CTC	30 CTC Leu
	CTTTCCGACC ACATTTCATT TCTTGCCTCG	90 CCGCCGCCGC C ATG Met	÷	GAG (Glu		CTC		CCC	280	GGC Gly	3 CTG Leu
0				CTC (180	CCC	230	GCC	· ·	ATG	AAG Lys
20)5533	80 3TTCG	130	CCT (-	CGC		TCC		GGC Gly	0 GAC ASP
	CGCGTCCGGG	CGTC		TCC (CTC		GCC Ala	270	ACC Thr	320 TAC GAC Tyr Asp
10		70 CG C		CCC '	0	GCT	220	ACC	2	ATC Ile	TAC
	3ACC(CTC	120	CGC (Arg	170	GCC		GCC		GTC Val	GCC Ala
	GGTCGACCCA	70 CGCTCCTCCG CCGTCGTTCG	Ä	CTC (GCC		GCT	260	GTC Val	310 GAC ASP
								•			

IGURE 6

	CAG Gln	450	CGG Arg		GCT Ala		AAG Lys	GTC Val		ATC Ile	069	CTG	
400	GGC G	4	GAC		AAG Lys		GAT Asp	O ACT Thr	640	AAG Lys	Ψ	GCG Ala	
	GCC Ala		AAC Asn		AAG Lys	540	AAG ATT Lys Ile	590 CTA AC Leu Th		CGG Arg		TCT	
	TTC	0,	AAG Lys	490	GGC Gly	u,	AAG Lys	GGC Gly		CAC His	089	GGG G1y	
390	AGG Arg	440	GGC Gly		GCC Ala		TCC	GGT Gly	630	GGT Gly	9	ATG Met	
m	ACC Thr		GAC Asp		GTC Val	530	CTC	580 ATG Met		AAA Lys		AAC Asn	
	CCC Pro		ATC Ile	480	ATT Ile	Ŋ	TCC	GGT Gly		GAG Glu		ACA Thr	9
380	TTC Phe	430	TAC	~	TGC		CAA	ACC Thr	620	ATC Ile	670	ATT Ile	FIGURE
38	AAA Lys		GGC Gly		TAC TYr		GGC Gly	570 GGA Gly	9	CTC		GCC Ala	FI
	TCC		ACG Thr	470	CGC	520	GCC	GTT Val		AAT		TAT	
	GCT	420	GCG	4	CTC		CTC	CTA		CAG Gln	099	CCA	
370	GAC		AAC		TGC		GAT Asp	60 GTG Val	610	GTT Val		ATT	
	TTC		TTC		GAC GAT ASP ASP	510	GCC Ala	56 GGA Gly		GGG G1y		rrc Phe	
	CGC Arg	410	GGC Gly	460			GAC Asp	GCC		GAC	029	TTT	
360	gac Asp	4	CGT		CTC		GAA Glu	AGG	009	* TCT Ser	9	CCG Pro	
,	ATC Ile		ATC Ile		CGG Arg	200	CTC	550 GAG Glu		TTC		TCC	

SUBSTITUTE SHEET (RULE 26)

		•										٠
	ACT Thr		ATC Ile	GCG		TCT	930	GAC Asp		GAG Glu	÷.	TAT
	TCA Z		CAT His		880	TTA Leu	6	AAG Lys		ATG Met		GAA Glu
	ATT	780	AAT Asn	830 GAG GCT Glu Ala		GCT		GAT Asp		GTT Val	1020	GCA
730	TCG ATT Ser Ile	7	GCC	ACT		AGG Arg	0	TGG Trp	970	TTG	.10	ATT Ile
•	TAT 'TYY		GCC	GGA Gly	870	TGC Cys	920	CCG Pro		GTA Val		ATT
	AAC '	0	GCT	820 GGA Gly	ω	GCC Ala		AGG		GGA Gly	. 01	CCG
720	CCA	770	TAT Tyr	GCT Ala		GTT Val		GCC TCA Ala Ser	096 *	GCT Ala	1010	GCG Ala
7	GGC Gly		TTT Phe	ATT Ile	0.5	TTC Phe	910		O1	$_{\rm GGG}$		GGA Gly
	ATG Met		TGC	10 ATG Met	098	$_{\rm GGA}$		ACT Thr		GAA Glu		CGG Arg
0	CTG	760	TAC TGC Tyr Cys	CTG Leu		GGA Gly		CAG Gln	950	$_{\rm GLY}^{\rm GGT}$	1000	AAA Lys
710	GGT Gly		AAC Asn	GAC Asp		TTA Leu	006	CCT Pro	9	ATG Met	.,	ATG Met
	TTG		TCC	0 GCT Ala	850	GGT Gly	O1	GAT Asp		GTG Val		GCA
	GAT Asp	750	ACT Thr	800 GAG GC' Glu Ala	٠	ATT Ile		GAT Asp		TTT Phe	066	CAT
700	ATC Ile	7	GCT Ala	GGT Gly		CCA Pro	890	AAT Asn	940	GGC Gly		GAG Glu
	GCC		TGT Cys	CGA	840	ATT Ile	8	AGG Arg		GAT Asp		TTG
	CTT	740	GCA Ala	790 CGC Arg	ω	GTC Val		CAA		CGT	980	AGC

SUBSTITUTE SHEET (RULE 26)

AGG Arg		GAT	1170	ACT Thr		GTT Val		ATC Ile	ATT Ile		AAT Asn
0 CCA Pro	1120	GAA Glu	11	GCG Ala		AAA Lys		ATG Met	1310 GCA ACC Ala Thr	1360	TTT Phe
1070 GAT CCA ASP Pro	⊣	CTC		CAT His		GCC ATT AAG Ala Ile Lys	1260	TCA Ser		, ,	CAA Gln
ACT Thr		AGT Ser	0.0	GCT Ala	1210	ATT Ile	12	AAG Lys	ATC Ile		AAT Asn
ATG Met	1110	AGC Ser	1160	AAT Asn		GCC Ala		ACT Thr	GCC Ala	1350	CCC AGC ATT Pro Ser Ile
060 CAT His	11	GAG Glu		ATA Ile		AAT Asn	20	GCA	1300 GAA Glu	, ,	AGC
1 TAT Tyr		ATT Ile		TAC Tyr	1200	ATA Ile	1250	AAT Asn	CTT		CCC
GCT Ala	00	TGC	1150	AAT Asn	ij	GAG Glu		ATC	GGT Gly	1340	CTT CAT Leu His
1050 TGT GAT Cys Asp	1100	TCG	 1	GTC Val		GCC Ala		GAA ATC AAA Glu Ile Lys	1290 SA GGA Sr Gly	13	
		TCC Ser		GAG Glu	90	CTT	1240	ATC Ile	T Se		TGG
AAC Asn		GTC Val	1140	GAA Glu	1190	GAT Asp			GCA		GGC
1040 GCA GTC Ala Val	1090	GGT Gly	H	CCT		$_{\rm GGG}$		AAG Lys	80 GGA Gly	1330	ACC Thr
1040 GCA G Ala Va		CTT Leu		TCA		GCT	1230	ACC Thr	1280 CTT GC Leu Gl		ACC Thr
GGT Gly		$\frac{\text{GGG}}{\text{Gl} Y}$	30	GTC Val	1180	CTT Leu	H	AAC Asn	TGT		ATA Ile
GGA Gly	1080	GCT GAT Ala Asp	113	GGG Gly		ACT Thr		AAG Lys	CAC His	1320	GGA Gly
1030 TTG Leu	1(GCT Ala		GCC Ala		TCT Ser	1220	TTC Phe	1270 GGA G1y	H	AAG Lys

FIGURE 6

1410	AAG CAG CAA Lys-Gln Gln		GGG CAC Gly His	1510	ATTCT ACTTGGTTCA	1570	TAAATGCCTT	1630	AGCCATTTAG	1690	CTCTGATTTA	1750	GTTATTTAAG		CT	
1400	AAC AAA AAG Asn Lys Lys	1450	GGA TTT GGA Gly Phe Gly	1500	TGA * * *	.1560	AGCAATTTTT	1620	GTCCTTTGAT AGTTCCTCGA	1680	ATCGAAGATG ATTCCCATTT TAAATCTAGT	.1740	TGTTGTCAAT	1800	ATCCAGCTTA	
06	ACT GTT GCC Thr Val Ala	1440	AAT TCT TTT Asn Ser Phe	1490	TTC AAG CCA Phe Lys Pro	1550	CAACTTGCAG	1610	GTCCTTTGAT	1670	ATTCCCATTT	1730	GTCATGTTTG	1790	GCTCTAGAGG	
1390	TTC AAC Phe Asn	1430	GCT ATC TCG A Ala Ile Ser A	1480	TCA GCT Ser Ala	1540	GATAGGGCTT	1600	GAATAGGTCG	1660	ATCGAAGATG	1720	AAGATTTTGT	1780	AAGGGCGGCC	
1380	TCG GTG GAC Ser Val Asp		AAC GTC Asn Val	1470	GTG GCA Val Ala	1530	CAGTTGCTGA	1590	CGTAATACCG	1650	ТАСТGТААТА	1710	AGACCAATGA	1770	ATAAAGCAAA AAAAAAAA AAGGGCGGCC GCTCTAGAGG	
1370	CCC GAG CCA Pro Glu Pro	1420	CAT GAA GTG His Glu Val	460 1.	AAC TCG GTT Asn Ser Val	1520	AAATGCACAC	1580	GTCGGAAGAG	1640	GATGATGTTT	1700	TGTATTAGAA	1760	ATAAAGCAAA	

FIGURE 6

Gln Cys Ala Pro Leu>

CCA CTA

TGC GCC

CGC CGC CGG ATT CTC TCC CAA

Arg Ile Leu

Arg Arg

Arg Arg Leu Ser

 1 CC

CGA CGC CTC

၁၅၁ Arg

9

180 120 AGAGAGAGGG ATCCATCGAA TGCGGCCACC CTCCTTTCAT CTTCGATTCA TTACCATACC ATTCCGCTGA TCCATTTTCC GCCTTTTCCG GGTCTTTCAT CCCAAAGGGT ATCCTTTTCT GTACGCCTGC AGGTACCGGT CCGGAATTCC CGGGTCGACC CACGCGTCCG CATAAAAGAG Pro> CCT CTC TGT ACG TGG CTC CTT GCC GCC TGC ATG TCT Pro Leu Cys Thr Trp Leu Leu Ala Ala Cys Met Ser> Met Pro Ala Ala Ser Ser> 330 TCC GAC CCT CTT CCG CCT TCC ATC TCC TCT CCT 190 200 210 220 230 ATCCTATCTT CTCAAAGGGT CAGTCAGTTC CCTCCA ATG CCT GCC GCC TCT TCC Ser Ser Phe His Pro Ser Asp Pro Leu Pro Pro Ser Ile Ser 170 110 370 50 320 270 160 100 40 360 310 260 150 30 90 350 300 250 140 20 TCC TTC CAC CCC TCC Leu Leu Ala Ser Sequence Range: 1 to 2369 CTC GCT 290 130 ACC Thr CTG

FIGURE

	GTC Val>	TCC Ser>	0.	CGG Arg>	57.0	CTG Leu>		CAG Gln>		CAT His>	ATA Ile>
	CTC	470 ACA Thr	520	CAC His		GCT Ala		AAA Lys	•	GGC G1y	710 GGC Gly
420	ACC CTC Thr Leu	4. TAT Tyr		AGG		GTG Val	0	ATC Ile	* 099	CTA Leu	AGT Ser
7	CAT A	TAC T		CGC	260	GCC Ala	610	AGT Ser		CCT Pro	ACG Thr
	5 × 01	r.	510	ACC (Thr	Ŋ	ATG		CCA Pro		ACT Thr	.0 GGA G1Y
0	AGT T Ser F	460 CAT GAC His Asp	u,	ACC 7		GCA Ala		AAG Lys	029	GTG Val	700 GAT GGA ASP GlY
410	TCC A	TGC C Cys F		CGC A	0	GAG (Glu	009	AAG Lys	9	GTG Val	CTT
	GGA 1 Gly S	CCC 1	200:	ATT (Ile i	550	AGG		AAG Lys		$\tt GGT\\ \tt G1Y$	CTG
	CGC G	450 GAG (Glu I	5	CCC /		TCC		ACA Thr	0	ATG Met	690 AAT Asn
400	CTC C	t TTC (Phe (AGA (Arg		CCT	290	$^{ m ACC}_{ m Th}_{ m T}$	640	GGA Gly	AAT Asn
	GCC C Ala I	TGC 1 Cys 1	0	TCC /	540	TCC	Ŋ	GTT Val		ACT	TAC
	TCC G	440 GCC 1 Ala (490	GGA G		GCT		GAA Glu		GTG Val	680 TTC Phe
390	TCC T	44 CTC (Leu A		TTC (Phe (CGA	0	CAG Gln	630	GTT Val	GTT Val
36	GCT T Ala 9	TAC (Tyr I		TTG .	530	AAT	580	GAA Glu		GTA Val	GAT Asp
	TCT G	ELA	480	* TCC ' Ser .	2	CTC		CCT		CGA Arg.	670 GAC CCT ASP Pro
380	CCT I	430 ACC TC Thr Se	4.	GCA 3		AGG (CAA	620	CGG	670 GAC CC ASP PJ

7. TGURE /

FIGURE 7

0.0	GCT Ala>	810	CTC Leu>		AAG Lys>		CTA Leu>	ATG Met>	00	AAG Lys>	1050	GCT Ala>
760	ATT Ile		AAG Lys		66C 61y		AAA GAG Lys Glu	950 GGA Gly	1000	AAG Lys		TCA
	AGA Arg		CCG Pro	0	GCT Ala	006		9 GGT Gly		$\mathtt{TAT}\\ \mathtt{Tyr}$		GGA Gly
	ACG Thr	800	GCC	850	ACC Thr		ATG Met	ATG Met		TCA	1040	AAT ATG Asn Met
750	CCT Pro	∞	GTG Val		CTG Leu		GTG Val	940 A GCA Er Ala	990	ATT Ile	1(AAT Asn
	TTT Phe		TGG Trp		ATG Met	068	GAT Asp	94 TCA Ser		AGG Arg		ACA Thr
	CAA Gln	0	$_{\rm GGT}$	840	TAC	ω	GAA Glu	GGC Gly		CTA Leu	30	ACC Thr
740	GCT	790	GAT Asp		CTA Leu		ACC Thr	ATT Ile	086	GCC	1030	GCT Ala
7	TGT Cys		ACA Thr		ATG Met	880	ATC Ile	930 CTC Leu	0,	GAA Glu		TTC
	GAT Asp		TCC Ser	830	TTC Phe	88	GGA Gly	GTT Val		ATT Ile		CCT Pro
0	ТТТ Phe	780	TTC Phe	ω	AAG Lys		$_{\rm G1y}^{\rm GGT}$	GGA Gly	970	GAT GCC Asp Ala	1020	TGT GTA Cys Val
730	ACC Thr		TCT Ser		GAC Asp		GAT Asp	920 TGC Cys	ģ	GAT		
	GAG Glu		AAG Lys	0	ATG Met	870	ACA Thr	9 AAA Lys		AAT Asn		TTT Phe
	ATA Ile	170	ATC Ile	820	AGG		TTA Leu	AGA Arg		TTC Phe	1010	CCC Pro
720	GAG Glu	7	GAG Glu		AAG Lys		GCA Ala	910 T AAA P Lys	096	GTA Val	1(AAT Asn
	AGC		GGA G1y		TCT	860	AAA Lys	91 GAT ASP		AAG Lys		ATG Met

FIGURE 7

	TCT Ser>		CAT His>	GCG. Ala>	01	TTG Leu>	1290	AGT Ser>		CTA Leu>	•	GAA Glu>
	ATA Ile		AAC Asn	1190 A GAT er Asp	1240	GCT	•	GAC		CTA Leu		GCA Ala
0	TCG	1140	GCG Ala	11 TCA Ser		CGA Arg		CCA TGG Pro Trp	0	CTA Leu	1380	TAC
1090	TAC ' Tyr	Т		66C G1y		TGC Cys	1280	CCA TGG Pro Trp	1330	GTG Val		ATT
	AAC '		AAT GCT Asn Ala		1230	GCA Ala	12	AGA Arg		GGA Gly		GCG ACT Ala Thr
	CCC	30	ATG	1180 TGC GGG Cys Gly	7	GTT Val		TCA Ser		GCT Ala	1370	GGT GCG Gly Ala
1080	GGG (1130	ATA	CTT		TTT Phe	0	GCT Ala	1320	${\tt GGA} \\ {\tt G1y}$	13	$_{\rm GLY}^{\rm GGT}$
Ţ	ATG (TGT	ATG Met	1220	GGT Gly	1270	AAA GCT Lys Ala	(1	GAA Glu		AGA
•	TGG	0	TTT Phe	170 GTG Val	12	GGA Gly		ACT		$_{ m GGG}$	0.5	
7 0 4	GGA Gly	1120	AAC Asn	1170 GAT GTG Asp Val		ATG Met		CCT	1310	ATG Met	1360	AAG Lys
1070	TTG (AGT	GCA Ala	0	GGT Gly	1260	GAC Asp	13	GTT Val		GCA Ala
	GAC		ACG	1160 3C GAA .Y Glu	1210	ATT Ile	-	TCC		TTT Phe		CAT
0	ATG	1110	GCA Ala	1160 GGC GAA Gly Glu		CCT		AAT Asn	0.0	GAT GGA Asp Gly	1350	GAG
1060	GCA	-	TGT Cys	AGA Arg		ATA Ile	1250	AGA Arg	1300	GAT Asp		TTG
	CTT		GCT Ala	ည္ ခ	1200	ATC Ile	12	CAG Gln		CGT Arg		GAG Glu
	ATG (1100	ACT	1150 ATA AT Ile I		GTA Val		TCC		AAT Asn	1340	GAG Glu

. ^		△		ω ά		C) M		رد در در در	A 1>		A u>
CCT Pro>	1480	GCT Ala>	1530	GCC Ala>		CAC His>		A ATG	A GTA c Val>	1720	GAA u Glu>
1430 C GAG or Glu	14	TTG		CAT His		ATC Ile		TCA	1670 GTT TCA Val Ser	ί	TTG
ACC Thr		GCT		GCC Ala	0	GCT CTT Ala Leu	1620	AAA Lys			AAT Asn
ATG Met			1520	AAT Asn	1570			ACC Thr	GCA Ala		CCG AAT ATT Pro Asn Ile
O CAC His	1470	GAG AAG Glu Lys	15	ATA Ile		CAA Gln		TCA	1660 GTG GAA Val Glu	1710	AAT Asn
1420 TAC CAC Tyr His	Ţ	ATA Ile		TAC		TAC Tyr	1610	AAT			CCG
GCC '		TGC	0	AAT Asn	1560	AAA GAG Lys Glu	16	GTT Val	GGT G1y		CAT His
GAT (1460	CTC	1510	GTA Val					GGT Gly	1700	TGG ATC Trp Ile
410 TGC Cys	14	ATT Ile		GAC Asp		ATC Ile	0	GAG TTA Glu Leu	1650 GCC Ala	, 	
ACT Thr		GTG Val		GAA Glu	1550	GAT Asp	1600		GCA Ala		$_{\rm GGG}$
TTC	0	GGA G1y	1500	AGG	15	GGA Gly		AGA Arg	GGA Gly	06	AGG ACT Arg Thr
00 AGT Ser	1450	GCT	←	TCT Ser		GCT Ala		AAC Asn	1640 rr crc eu Leu	1690	
1400 GGG AGT Gly Ser		GGA Gly		GTC Val	0	CCG Pro	1590	CAA Gln	υğ		ATA Ile
GGT (GAT Asp	1490	GGA Gly	1540	ACT Thr	-	GGC Gly	CAC		GCA Ala
	1440	CCT Pro	14	TCA Ser		TCC		TTC Phe	1630 ATT GGT Ile Gly	1680	cAG Gln
1390 TTT CTA Phe Leu	H	CAC		CAG Gln		ACA Thr	1580	TGT	1630 ATT GC Ile G	-1	GTT Val

FIGURE 7 5/7

FIGURE 7

1770	AAG AAG Lys Lys>		TTT GGT Phe Gly>	1870	GTTTCCGTGT	1930	GTTGGTAGCT	1990	GAACCATGAC	2050	GTAGAGCAAT	2110	GTTGTACTTT	2170	CACGTAGTAA
1760	GTG GGT CCT Val Gly Pro	1810	TCA TTT GGG Ser Phe Gly	1860	ATC TAG GAC Ile ***>	1920	ACTCCAGCAT	1980	CCATGAGTTT TGTGTCCGGA GCTTTAGTCG	2040	ACTCCTTGCT AGAATTGTTG	2100	CCTTGCAATA	2160	CGAGCTTTTC ATCGAGTCAG TGAAGAAGAG AACAAAGCTG TTAACTCGGG
1750	AAA TTG CTC Lys Leu Leu	1800	TTG TCT AAT Leu Ser Asn	1850	GCC CCT TAC Ala Pro Tyr	1910	AGTTTTGAGG	1970	TGTGTCCGGA	2030	ACTCCTTGCT	2090	AAATCTCCCT	2150	AACAAAGCTG
	GAT ACA Asp Thr	1790	GTC GGT Val Gly	1840	CTC TTC Leu Phe	1900	TCAAAGCTGA	1960	CCATGAGTTT	2020	CACTTGATAT	2080	TTTTTCTCTG	2140	TGAAGAAGAG
1740	GAA GGC GTG Glu Glu Gly Val		AAC GTT AAG Asn Val Lys	1830	TCG TCC ATA Ser Ser Ile	1890	GTGGAATTCT ACTCAACATA TCAAAGCTGA AGTTTTGAGG	1950	CTAGACATGC	2010	CTCATGGCGA CACTTGATAT	2070	TCATATTTT	2130	ATCGAGTCAG
1730	AAC CCA GAT GAA Asn Pro Asp Glu	1780	GAG AGA CTG Glu Arg Leu	1820	GGG CAC AAC Gly His Asn	1880	GTGGAATTCT	1940	CCTTACGTCT	2000	GGATTGAGTA	2060	ATTCATTATC	2120	CGAGCTTTTC

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2230	AATTTGTAA	2290	ATGTATGTTT	2350	AAAAAAAA
2220	CCATITIGCCC TITIGITITICS TCICIATITIC ATCACCGITIT IGIGGITITIA AAAITIGIAA	2280	AACTAGAAGA CTGGTTTAGA TTGGTTTGTT TTCTCATTGA TAATTGGGGR ATGTATGTTT	2340	TGGAAATAAA AAAAAAAA AAAAAAAA AAAAAAAA AAAAAA
2210	ATCACCGTTT	2270	TTCTCATTGA	2330	AAAAAAAAA
2200	TCTCTATTTC	2260	TTGGTTTGTT	2320	AAAAAAAAA
2190	TTTGTTTTGC	2250	CTGGTTTAGA	2310	AAAAAAAAA
2180	CCATTTGCCC	2240	AACTAGAAGA	2300	TGGAAATAAA

2360 AGGGGGCG CTCTAGAGG

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) *)	CACACCAAAC	120	AC	180	ပ္	0 *	G	0 *	F	0 *	₽	0 *	\circ	0 *	Ø
	CACAC		ACAGACAGAC	18	TCTTCGATTC	240	TCCCAAAGGG	300	CCTGCCGCCT	360	TCTACCTCCT	420	CTCTCCCGCC	480	CGCGGATCC
00	GACGCCAACC	. 110	AGACAGACAG	170	CCTCCTTTCA	230	GGGTCTTTCA	290	CCCTCCAATG	350		410	TCGCCGACGC	470	CTICTGCTIC CTCCGCCCTC CGCGGATCCA
7 N	ACGCGTCCGC	. 100	CATTGGCAGC	160	ATGCGGCCAC	220	CGCCTTTTCC	280	TCAGTCAGTT	340		400	TCTCCTCTCC	460	CTTCTGCTTC
30	GGGTCGACCC	06		150	GATCCATCGA	21.0	ATCCATTTTC	270	TCTCAAAGGG	330		390	CCGCCTTCCA	450	GCCCCACTAC
20	CGGAATTCCC	08	TCTCTTCTCA	140	GAGAGAGAGG	200	CATTCCGCTG	. 260	TATCCTATCT	320		380	CGACCCTCTT	440	CTCCCAATGC
10	-A-CNTGGTC	. 10	TTCCTCAGCT	130	CCATAAAAGA	190	ATTACCATAC	250	TATCCTTTTC	310	CTTCCCTGCT	370	TCCACCCTC	430	GCCGGATTCT
		CGGAATTCCC GGGTCGACCC ACGCGTCCGC GACGCCAACC	10 20 30 30 30 30 30 30 30 30 30 30 30 30 30							9	9				

FIGURE 8 1/5

ω

540	GACTACTATA	* 009	CGGAGGCTCA	099	ACAGGAAGTT	720	TTGTGACTGG AATGGGTGTG	780	ATGGAACGAG	840	TTGCTGGAGA	* 006	GGATGGACAA	* 096	GAATCACCGA	1020	CAGCAATGGG
530	GCCCTGCCAT	290	CCGCAGGCAC	650	TGCAACCTGA	710		770	AATCTGCTTG	830	CCTACGAGAA	890	CTCTCTAAGA	950	ACAGATGGTG	1010	CTCATTGGCT
520	CCTGCTTCGA GCCCTGCCAT GACTACTATA	580	TTCGCACCAC	640	GCCGTGGCTC	700	CGGCGAGTAG	760	TTTCTACAAT	820	TGCTCAATTT.	880	GGCCCCGAAG	940	TACATGCTGA CTGCTGGCAA GAAAGCATTA ACAGATGGTG	1000	ATAAAAGAAA ATGCGGAGTT CTCATTGGCT
510	TCTTACCTCG	570	TCCAGACCCA	630	GGAGGCAATG	069	TATCAAACAG	750	TAGGCCATGA ACCTGATGTT	810	CCTTTGATTG	870	ATGGTTGGGT	930	CTGCTGGCAA	066	
200	CCTCGTCACC	260	CTTGTTCGGA TCCAGACCCA	620	CCCTTCCAGG GGAGGCAATG GCCGTGGCTC TGCAACCTGA ACAGGAAGTT	089	AGAAGCCAAG	740		800	GAGATAGAGA	860	GATCAAGTCT TTCTCCACAG ATGGTTGGGT GGCCCCGAAG	920		086	AAAGAGCTAG
490	GTTTCCATAC	550	CATCCGCATC	610	ATCGAGCTTC	0.49	ACCACAAAGA	730	GTGACTCCTC	790	TGGCATAAGC	850	GATCAAGTCT	910	GTTCATGCTA	970	AGATGTGATG

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TACGCAGAAT TICTAGGIGG GAGITICACT IGCGAIGCCI ACCACAIGAC	TGCGATGCCT	GAGTTTCACT	TTCTAGGTGG	TACGCAGAAT	TGCGACTATT
1500	1490	1480	1470	1460	1450
AGAAAAGAGG	GAGCATGCAA	AGAGGAGTTG	TGCTACTACT	GGAGCTGGAG	TATGGGGGAA
1440	1430	1420	1410	1400	1390
ATGGATTTGT	AGTAATCGTG	ACCATGGGAC	AAGCTTCAAG	GAGAAATTCC GACCCTACTA AAGCTTCAAG ACCATGGGAC AGTAATCGTG	GAGAAATTCC
1380	1370	1360	1350	1340	1330
CTTTGTCCCA	GCATGCCGAG	ATCATACCTA TTGGTATGGG AGGTTTTGTT GCATGCCGAG	TTGGTATGGG	ATCATACCTA	AGATGCGGTA
1320	.1310	1300	1290	1280	1270
GCGGGGGCTC	GTGATGCTTT GCGGGGCTC	CGAAGCAGAT	GCGAACCATA TAATCAGAGG		AATGAATGCT
1260	1250	1240	1230	1220	1210
ACTTTTGTAT	GCAACGAGTA ACTTTTGTAT	GGGCCCAACT ACTCGATATC TACTGCTTGT	ACTCGATATC		GGGATGGATG
1200	1190	1180	1170	1160	1150
CAATGGACTT	GCTATGCTTG	TATGGGATCA	CTACCACAAA	GTACCTTTCG	TCCCTTTTGT
1140	1130	1120	1110	1100	1090
AGAAGATGAA	ATTTCATATA	AGCCCTAAGG	ATGCCATTGA	TGGAATGAAG GTATTCAATG ATGCCATTGA AGCCCTAAGG ATTTCATATA	TGGAATGAAG
1080	1070	1060	1050	1040	1030

FIGURE 8

1560	TGGCTCAGTC	1620	CICCGGCTGG	1680	AGTTAAAAGT	1740	TGGAAGCAGT	1800	TGGAAAACCC	1860	TGAACGTTAA	1920	TCTTCGCCCC	1980	CTACTCAACA TATCAAAGCT GAAGTTTTGA	2040	CTCTAGACAT GCCCATGAGT TTTGTGTCCG
1550	GAGAAGGCTT	1610	GCCACATCCA	1670	CAAAACAGAG	1730	TCTCGGAGCA GCCGGTGGTG	1790	AATATTAATT	1.850	GGGTCCTAAG AAGGAGAGAC TGAACGTTAA	1910	TCGTCCATAC	1970	TATCAAAGCT	2030	GCCCATGAGT
1540	TCTCTGCATA	1600		1660	CTCTTATCCA CTGTTTCGGC CAAAACAGAG AGTTAAAAGT	1720	TCTCGGAGCA	1780	GATCCATCCG	1840	GGGTCCTAAG	1900	TGGGCACAAC	1960		2020	
1530	CTGGAGTGAT	1590	TAAATTACAT AAATGCCCAT	1650	CTCTTATCCA	1710	TTGGTCACCT	1770	GGACTGGGTG	1830	GTGGATACAA AATTGCTCGT	1890	TTGGGTTTGG	1950	GACGTTTCGT GTGTGGAATT	2010	CTCCTTACGT
1520	CCTGATGGAG	1580	AGGAGTCTCT AGGGAAGACG	1640	GAGTACCAAG	1700	AAATCAATGA	1760	CAGGCAATAA	1820		1880	TCTAATTCAT	1940		2000	ATGTTGGTAG
1510	CGAGCCTCAC CCTGATGGAG	1570	AGGAGTCTCT	1630	AGATATCAAA	1690	TAATTCAACC	1750	TTCAGTAGTT	1810	AGATGAAGGC	1870	GGTCGGTTTG	1930	TTACATCTAG	1990	GGACTCCAGC

FIGURE 8

		ATCC	2370 GCTCTAGAGG		2350 2350 2350
TTTTCTCAAA	GATTGGTTTG	GACTGGTTTA	TAAAATTTGT AAAACTAGAA GACTGGTTTA GATTGGTTTG		TTTGTGGTTT
2340	2330	2320	2310	2300	2290
TCATCACCGT	GCTCTCTATT	CCTTTGTTTT	TGTTAACTCG GGCACGTAGT AACCATTTGC CCTTTGTTTT	GGCACGTAGT	AACTCG
2280	2270	2260	2250	2240	2230
AGAACAAAGC	AGTGAAGAAG	TCATCGAGTC	CTCCTTGCAA TAGTTGTACT TTCGAGCTTT TCATCGAGTC	TAGTTGTACT	rtgcaa
2220	2210	2200	2190	2180	2170
TGAAATCTCC	TTTTTTCTC	TCTCATATTT	CTAGAATTGT TGGTAGAGCA ATATTCATTA TCTCATATTT	TGGTAGAGCA	AATTGT
2160	2150	2140	2130	2120	2110
ATACTCCTTG	GACACTTGAT	TACTCATGGC	CGGAACCATG ACGGATTGAG TACTCATGGC GACACTTGAT ATACTCCTTG	CGGAACCATG	GAGCTTTAGT
2100	2090	2080	2070	2060	2.050

8 3X0514 5/5

Sequence Range: 1 to 1580

GGG G1y>	0	TCG Ser>	150	GTG Val>		GAT Asp>		TCT Ser>	AAA Lys>	340	CGC Arg>
50 TCT (Ser (100	CAT		AGG Arg		GGT Gly		GGA Gly	290 GCT Ala	3,	ATC Ile
GCA Ala		CAG Gln		AAA Lys	0	TTG	240	ATT Ile	2 CTT Leu		GGG Gly
AAT (Asn i		ACT Thr	140	TCC	190	TCT Ser		TTA Leu	GAT Asp		ACG
2G 1a	90	GCA	7	GTC Val		CAG Gln		AAA Lys	280 AAT GAT Asn Asp	330	CGA Arg
40 ATG G		AGG		TTT Phe		AGG Arg	230	TGC Cys	28 AAT Asn		GTC Val
GCTGGG		AGA	0	GAG Glu	180	GAC	N	GGA Gly	TCA Ser		ACT Thr
GCT	80	CTG	130	TCG		TCT		AGA Arg	GTC Val	320	ATT Ile
30 CGTT		GCC		TCC		GAT Asp	0	AGT Ser	270 CAA Gln	.,	TGG Trp
GTTT		CCT		TCT Ser	170	CAG Gln	220	GTG Val	CTT Leu		GAA Glu
o A GA	70	GTT Val	120	GGA Gly	-	GTT Val		CTT Leu	GCT	310	GAT Asp
20 agaga	7	TCA		CGT Arg		GCC Ala		AGG Arg	260 CCA Pro	3.	AAT
TTCA		TCT		TCT	0	AGT Ser	210	CCG Pro	ATA Ile		ACC Thr
10 GG A'		GGT Gly	110	TCG	160	TGT Cys		TCG Ser	GCT Ala		GAC
AATC	09	CTG	7	TCA		TGC Cys		CGC Arg	250 GGT TCT Gly Ser	300	GTC Val
10 CCTGAATCGG ATTCAAGAGA GAGTTTCGTT		TTT		ATT Ile		TTT Phe	200	TCT	250 GGT TC Gly Se		ATT Ile

IGURE

390	TCA Ser>		GAT Asp>		GGC Gly>	TTG Leu>	280	GTC Val>	630	GTG Val>		GGA. Gly>
, 1	GCA A		AAT Asn		TTC Phe	530 CCT Pro	28	TTA Leu		CTA Leu		CGG Arg
	TTA (Leu 1	0	GCA	480	CTT Leu	5 AAT Asn		GGT Gly		ATT Ile	019	GAT Asp
380	AAT ?	430	GAC (Asp		GAC Asp	AAG Lys		TTG Leu	620	AAT Asn	9	ACC Thr
38	ACA 7		GTA (Val		7D =	A 10	570	GTG Val	9	AAC Asn		TGG Trp
	CTT /		CAG (Glu	470	CCT Pro	520 TGC AAA Cys Lys		TTT Phe		TTT Phe		GAC
0	AGT (420	GCA	· 4	ACC Thr	GGC Gly		GGA G1y	610	$_{\rm GLY}^{\rm GGT}$	099 *	GTT Val
370	GAT Asp	-	ATG		TCT Ser	CTT	260	AGT Ser	61	GGG		\mathtt{TAT}
•	AAA (Lys		GAG Glu	0	ACT Thr	510 GCA Ala	Ľ	TGC Cys		GGT Gly		CGG Arg
	GGT	410	CTA	460	TGT Cys	AAA Lys		GCA Ala		AGA Arg	650	CTT TCT Leu Ser
360	TCA	4	GCT		ATG Met	TCG Ser	550	GCT	009	ATT Ile	Ū	
	CTC Leu		AAA Lys		TTG	500 ATA Ile	55	ACC Thr		CAC His		TCT Ser
	GTT Val	0	AGG	450	GTT Val	5 CAG Gln		ATT Ile		TGC	640	GAT Asp
350	AGG Arg	400	GCA		ATG Met	CCT	÷	GAC Asp	590	GCT Ala	9	GCT
Ж	CGA		GCA Ala		GAT Asp	O GCT Ala	540	TAC Tyr	.,	GCT Ala		GGT Gly
	AAC Asn		GAG Glu	440	GTG Val	490 AGT G		Ser		TCA		ATT Ile

FIGURE 9
2/5

Ile Glu Ser Ala Leu Gly Lys> Ile Asp Trp Leu Leu Leu His Gln Ala> GGA GAT GGG CAA AGG CAT CTA AAA GCT GCA ATC AAA GAA GAT GAA GTT Gly Asp Gly Gln Arg His Leu Lys Ala Ala Ile Lys Glu Asp Glu Val> Glu Val Phe Arg> Ile Arg Asp Phe Pro Pro Arg> 1010 TCC AAC ATC GAC TGG TTG CTG CTT CAT CAG GCA Leu Phe Ala Phe Asp Leu His Ser Asp> GTG CAG TCA Val Gln Ser> 870 CTT GGA AAG CCA AGG GAG GTA TTC CGC TTT GAT TTG CAT AGC GAT GGA CAT AAT GGG TCC ATC AGA GAT TTT CCA 720 910 CCT CAG TCA ATC GAA TCA GCA Gly Ala Val Val GCT GTA GTG 860 TGC ATC CAA ATG AAC GGT AAA Cys Ile Gln Met Asn Gly Lys 810 950 710 GCT GGA 006 850 Pro Gln Ser 750 CTC TTT (GAT AAA GCC CTG GGA CAT AAT GGG TCC Asp Lys Ala Leu Gly His Asn Gly Ser Asp Ala Ala 800 GCT940 700 Ser Asn Glu Glu Asp Gly GAG GAA GAT GGG GAT 890 CGC TCT GTG C Arg Ser Val F Thr Cys Ile Leu Phe Gly GGA 840 790 TCTSer GCC GGT CTT AAT GGA Ala Gly Leu Asn Gly TTT TAC 930 CTC 069 Ser Cys TCT TCA TTT GCT TGC TGT GAT GCT Cys Asp Ala TGT ATT Ala Arg Ser 780 970 Phe ACA ' 920 680

FIGURE 9

CCT CAA Pro Gln>	1110	GCG GCA Ala Ala>		GTG AAG Val Lys>		ACA TGG Thr Trp>	1260	CACTGCAGCT	1320	AAGAAGTCAG	1380	TCGTTCCCCT	
TA GAG GTT eu Glu Val	1100	ACT Thr	1150	AGT GGA AAT Ser Gly Asn	1200	GGA Gly	1250		1310	CCANAAAAAG	1370	TTGCCCTTTT	
ACA CGT Thr Arg	060	TAC GGG Tyr Gly	1140	GTG AGG Val Arg	1190	TTT GGC Phe Gly	1240	AA GACTGAA **>	1300	SCTTCCATGA	1360	CTTCATCACA	
GTA Val	\leftarrow	TTG GCA Leu Ala	30	GAC GAA Asp Glu	1180	ACC GCA Thr Ala	1230	TGG GGA Trp Gly	1290	CGAAATTTT (1350	CGACACGAT	
ATT GAT Ile Asp	1080	TCA AAC Ser Asn	11	GCA CTA Ala Leu	7.0	3CA Ala	1220	TT ATC AGG le Ile Arg	1280	GATGTTTC A	1340	CAAGCAAC A	
* AAT CAG AGG AT Asn Gln Arg Il	1070	GAA CGA ATT A1 Glu Arg Ile Il	1120	TCC ATT CCC TS		CCG GGT CAC G' Pro Gly His V	1210	GGT TCT GCT A Gly Ser Ala I	1270	TCCTCTCAAA CC	1330	TCTTTTATGG AG	
	* CAG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT Gln Arg Ile Ile Asp Ala Val Ala Thr Arg Leu Glu Val	CAG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT Gln Arg Ile Ile Asp Ala Val Ala Thr Arg Leu Glu Val 1070 1090 1100	CAG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT CCT Gln Arg ile ile Asp Ala Val Ala Thr Arg Leu Glu Val Pro 1070 1080 1100 1100 1 1090 1100 1 100	CAG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT CCT GIN Arg Ile Ile Asp Ala Val Ala Thr Arg Leu Glu Val Pro 1070 1080 1100 1100 1100 1 1070 1 1080 1130 1140 1150	CAG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT CCT CGIN Arg Ile Ile Asp Ala Val Ala Thr Arg Leu Glu Val Pro CGA ATT ATC TCA AAC TTG GCA AAT TAC GGG AAC ACT AGT GCG AAC ATT Ser Ala Arg Ile Ile Ser Asn Leu Ala Asn Tyr Gly Asn Thr Ser Ala ATT CCC TTG GCA CTA GAC GAA GCT GTG AGG AAC GGA AAT GTG Ile Ile Ser Asn Leu Ala Asn Tyr Gly Asn Thr Ser Ala ATT CCC TTG GCA CTA GAC GAA GCT GTG AGG AGT GGA AAT GTG Ile Pro Leu Ala Leu Asp Glu Ala Val Arg Ser Gly Asn Val	CAG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT CCT CGIN Arg Ile Ile Asp Ala Val Ala Thr Arg Leu Glu Val Pro CGA ATT ATC TCA AAC TTG GCA AAT TAC GGG AAC ACT AGT GCG AAC AIS AIB ASN TYR GIY ASN Thr Ser Ala AII CC TTG GCA CTA GAC GAA GCT GTG AGG AGT GGA AAT GTG Ile Pro Leu Ala Leu Asp Glu Ala Val Arg Ser Gly Asn Val III Pro Leu Ala Leu Asp Glu Ala Val Arg Ser Gly Asn Val 1170	CAG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT CCT CI GIN Arg Ile Ile Asp Ala Val Ala Thr Arg Leu Glu Val Pro CI CGA ATT ATC TCA AAC TTG GCA AAT TAC GGG AAC ACT AGT GCG CAAC ACT CAACA ACT GCA AAT GCG CAAC ACT GCG CAAC ACT CAACA ACT GCA ACT GCA AAT GCG CAAC ACT GCG CAAC ACT ACT ACT ACT ACT ACT ACT ACT AC	CAG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT CCT CAG GGIn Arg Ile Ile Asp Ala Val Ala Thr Arg Leu Glu Val Pro Glu CIn Arg Ile Ile Ser Asn Leu Ala Asn Tyr Gly Asn Thr Ser Ala And Thr Ser Ala Ash CT CC TTG GCA CTA GCT GCG ACT GCG GCA ACT CC TTG GCA CTA GCT GCG ACT GCG GCA ACT CC TTG GCA CTA GCT GCA ACT Thr Ser Ala And Thr CCC TTG GCA CTA GCT GCA ACT GCG GCT GCG GCT GCG ACT GCG GCT GCG ACT ACT THE ACT GCG ACT GCG GCC GCG GCC GCG CCC GCG CCC GCG ACT ACT THE ACT ACT GCG ACT GCG ACT ACT THE ACT	CAG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT CCT CR GIN Arg Ile Ile Asp Ala Val Ala Thr Arg Leu Glu Val Pro GI 1070 1080 1090 1100 1110 1120 1130 1140 1150 1150 1170 1180 1180 1190 1190 1200 1	CAG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT CCT CP GIN Arg Ile Ile Asp Ala Val Ala Thr Arg Leu Glu Val Pro GI 1070 1080 1090 1110 1120 1130 1140 1150 1180 1190 1190 1150 1117 1180 1190 1200 1200 1200 1200 1200 1200 1200 1200 1220 1230 1230 1230 1310 1310 1310 1310 1310 1310 1310 1310 1310	CGG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT CCT CP GIL AND AND AID AND Thr AND Leu GIU VAI Pro GIL AND AND THE SET AND Leu AID AND THY GIY AND THY GOG GCG GCG GCG GCG GCG ATT CCC TTG GCA CTA GCA GCA GCA GCA GCA GCA GCA GCA GCA GC	CGA AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT CCT CAGG AND 110 1080 11070 1080 11080 11090 1100 1111 CGA ATT ATC TCA AAC TTG GCA AAT TAC GGG AAC ACT AGG GG GG ACG TIC CAGG ATT ACG CCA AAT TAC GGG AAC ACT AGG GG GG ACG TIC AGG AACG AGG AGG AGG AGG AGG AGG AGG A	CAG AGG ATC ATT GAT GCA GTA GCA ACA CGT CTA GAG GTT CCT CA CIL GIN Arg Leu Glu Val Pro GIN Arg Ile Ile Asp Ala Val Ala Thr Arg Leu Glu Val Pro GIN Arg Ile Ile Ser Asn Leu Ala Asn Tyr GIY Asn Thr Ser Ala Ala Ala And Tyr GIY Asn Thr Ser Ala Ala Ala And Tyr GIY Asn Thr Ser Ala Ala Ala CGA ATT CC TTG GCA CTA GAC GAA GCT GGG AAC ACT AGT GGG GG GC TIS GCA CTA GAC GAA GCT GGA AAT GGG AC GCT GGG AAC ACT AGT GAA GCT GCA GCT GGA AAT GCG GCT GGG AAC GCT GGG AAC GCT AGT GGG GCT GCT AGT GCA ATT GCC GCA GCT GGA AAT GCA AGT GCA ATT GCC GCG GCC GGA CTC ACA TY GCT GCT ATT ATC ACG ACC GGA TTT GCC GCG GCC GGA CTC ACA TY GCT GCT ATT ATC AGG GGG TAAA GACTGAA GCCGAGCCAG CACTGCA GCT ATT ATC AGG GGG TAAA GACTGAA GCCGAGCCAG CACTGCA TTTTTATGG AGCAGCAAC ACGACACGAT CTTCATCATCAC CANAAAAAG AAGAAGTTTT TCGTTTTTTTTTTTTTTTTTTT

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				1570 1580 AAAAAAAA AAAAAAAAA	1570 AAAAAAAAA
ааааааааа	AAAAAAAAA	TTTGCTAAAA	GAGATGACAG CATAAACATC ATGTTTATAT TTTGCTAAAA AAAAAAAAA AAAAAAAAA	CATAAACATC	GAGATGACAG
1560	1550	1540	1530	1520	1510
CGGGACATTG	CATTTTGTCT	GCTTTTACTT	TAAGTTATTT GTTTCTTGTT TAATTGTTCA GCTTTTACTT CATTTTGTCT CGGGACATTG	GTTTCTTGTT	TAAGTTATT
1500	1490	1480	1470	1460	1450
TTGTCCCCAA	ATAGTTTCTT	TACAATACCC	TTTCCATTAG TTTGATGATT TTGCTGACAA TACAATACCC ATAGTTTCTT TTGTCCCCAA	TTTGATGATT	TTTCCATTAG
1440	1430	1420	1410	1400	1390

FIGURE 9 5/5

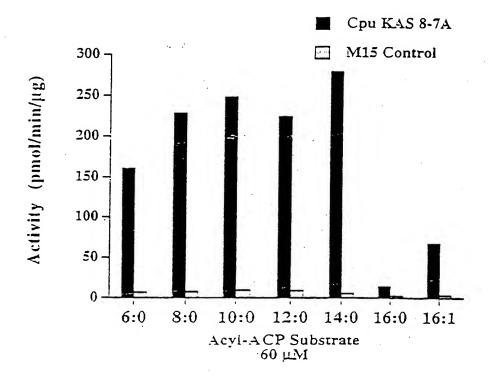


FIGURE 10

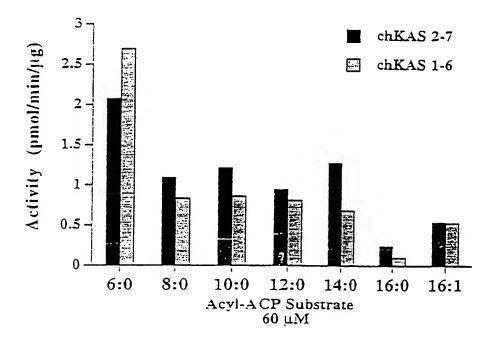


FIGURE 11

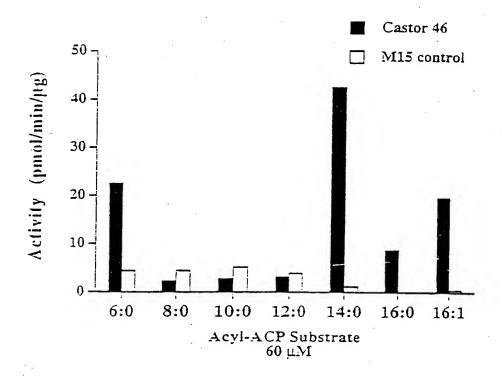
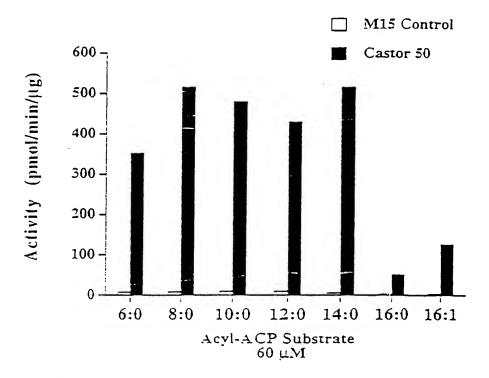
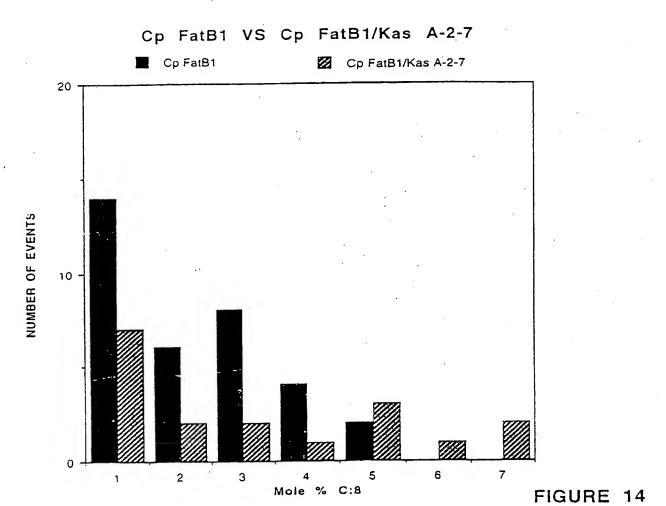


FIGURE 12



E328013-28

FIGURE 13



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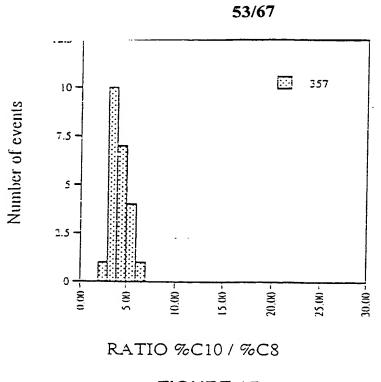


FIGURE 15

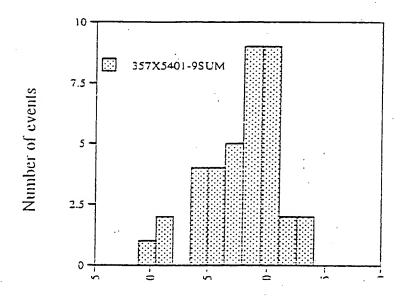
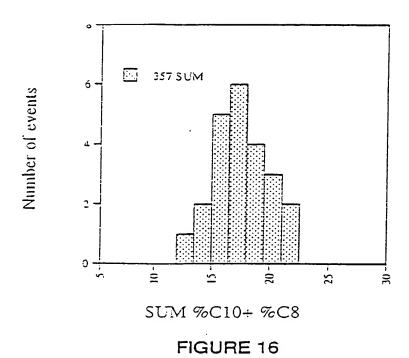


FIGURE 15 2/2



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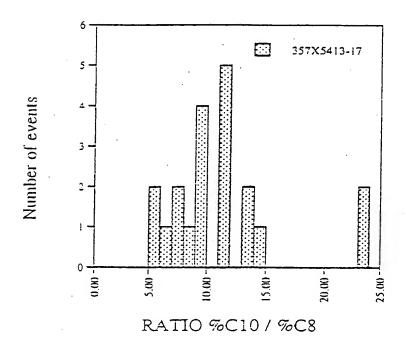


FIGURE 17

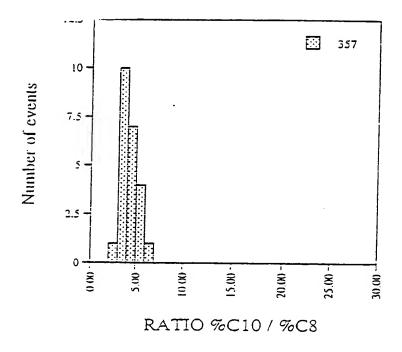


FIGURE 17

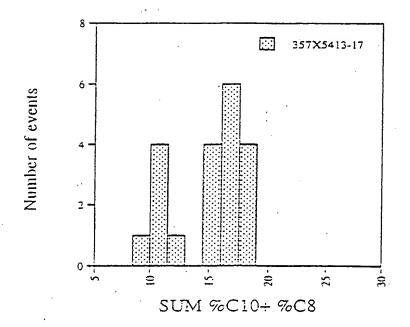
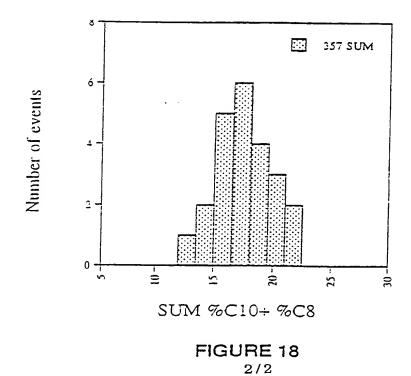
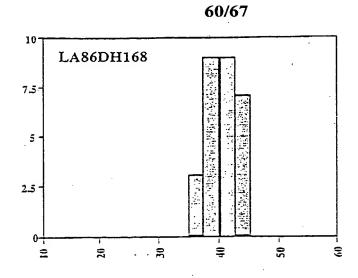


FIGURE 18 1/2

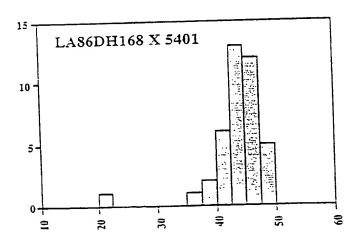






12:0 levels (w%)

FIGURE 19



12:0 levels (w%)

FIGURE 19 2/3

Number of independent events

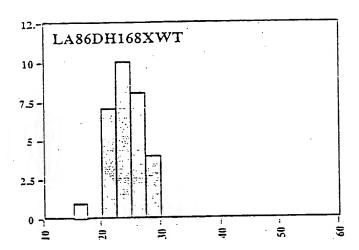


FIGURE 19 3/3

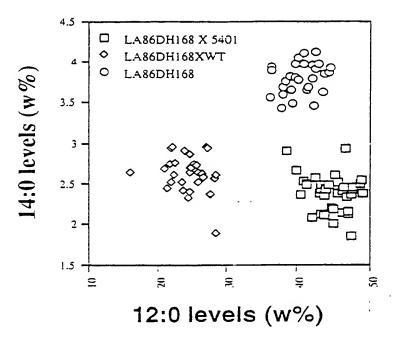
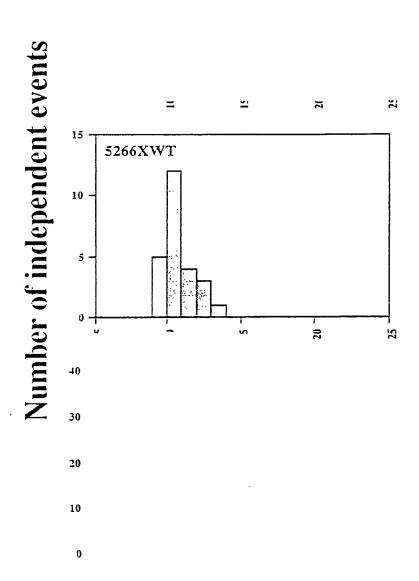


FIGURE 20

Number of independent events

18:0 levels (w%)

AFIGURE ~21



18:0 levels (w%)

FIGURE 21 2/3

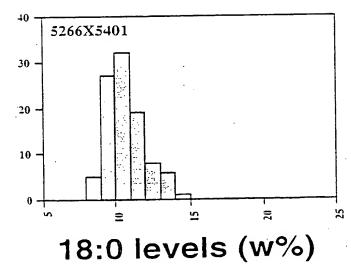


FIGURE 21 3/3

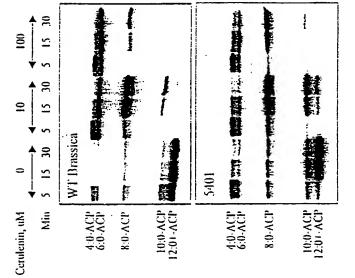


FIGURE 22

inte. ..ional Application No PCT/US 98/07114

		1	90/0/114
A. CLASSI IPC 6	FICATION OF SUBJECT MATTER C12N15/82 C12N15/54		
According to	o International Patent Classification(IPC) or to both national classif	ication and IPC	
B. FIELDS	SEARCHED		
Minimum do IPC 6	cumentation searched (classification system followed by classifica ${\tt C12N}$	tion symbols)	
Documentat	tion searched other than minimumdocumentation to the extent that	such documents are included in the fie	lds searched
Electronic da	ata base consulted during the international search (name of data b	ease and, where practical, search terms	s used)
C DOCUME	ENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the re	plevant nassages	Relevant to claim No.
Calogory	Ondation of document, with indication, where appropriate, of the fe	siovani passayos	Train value to cidim No.
X	WO 95 06740 A (MAX PLANCK GESELL;TOEPFER REINHARD (DE); MARTINI (D) 9 March 1995 see page 16, paragraph 1; claim	NORBERT	15,22
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"A" docume conside "E" earlier di filing da "L" docume which is citation "O" docume other m"P" docume later th	nt which may throw doubts on priority claim(s) or so cited to establish the publicationdate of another a or other special reason (as specified) and the repeated to a constant of the repeated of an oral disclosure, use, exhibition or neans and prior to the international filing date but an the priority date claimed	"T" later document published after the or priority date and not in conflicted to understand the principle invention. "X" document of particular relevance cannot be considered novel or involve an inventive step when "Y" document of particular relevance cannot be considered to involve document is combined with one ments, such combination being in the art. "8" document member of the same properties.	ct with the application but a or theory underlying the strength of the cannot be considered to the document is taken alone as inventive step when the or more other such docupobious to a person skilled patent family
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	nailing address of the ISA	Authorized officer	
	European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Maddox, A	

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Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This Inte	ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1.	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. X	Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically: see FURTHER INFORMATION sheet
з. 🗌	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking(Continuation of item 2 of first sheet)
This Inte	ernational Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4.	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark	on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Claims Nos.: 1-14,19,20,21,26,27,28

Remark : Claims 1-14 were not provided to the ISA at the time of search and hence the subject matter of these claims and the dependent claims 19,20,21,26,27,28, could not be defined.

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